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TUMOR NECROSIS FACTOR (TNF) INHIBITOR AND METHOD FOR OBTAINING THE SAME

Cross-Reference to Related Application

This is a divisional of 08/346242, filed 11/28 and issued 11/7/00 as 6,143,866, which is continuation-in-part of co-pending application Serial No. 07/479,661 filed February 7, 1990, which is in turn a continuation-in-part of applications Serial Nos. 07/381,080 filed July 18, 1989, and 07/450,329 filed December 11, 1989 for "Tumor Necrosis Factor (TNF) Inhibitor and Method for Obtaining the Same," both abandoned.

BACKGROUND OF THE INVENTION

Tumor necrosis factors are a class of proteins produced by numerous cell-types, including monocytes and macrophages. At least two TNFs have been previously described, specifically TNF alpha and TNF beta (lymphotoxin).

These known TNFs have important physiological effects on a number of different target cells involved in the inflammatory response. The proteins cause both fibroblasts and synovial cells to secrete latent collagenase and prostaglandin E2, and cause osteoblastic cells to carry out bone resorption. These proteins increase the surface adhesive properties of endothelial cells for neutrophils. They also cause endothelial cells to secrete coagulant activity and reduce their ability to lyse clots. In addition they redirect the activity of adipocytes away from the storage of lipids by inhibiting expression of the enzyme lipoprotein lipase. TNFs cause hepatocytes to synthesize a class of proteins know as "acute phase reactants" and they act on the hypothalamus as pyrogens. Through these activities, it has been seen that TNFs play an important part in an organism's response to stress, to infection, and to injury. See, e.g., articles by P.J. Selby et al. in Lancet, Feb. 27, 1988, pg. 483; H.F. Starnes, Jr. et al. in J. Clin. Invest. 82: 1321 (1988); A. Oliff et al. in

Cell 50:555 (1987); and A. Waage et al. in Lancet, Feb. 14, 1987, pg. 355.

However, despite their normally beneficial effects, circumstances have come to light in which the actions of TNFs are harmful. For example, TNF alpha injected into animals gives rise to the symptoms of septic shock; endogenous TNF levels have been observed to increase following injection of bacteria or bacterial cell walls. TNFs also cause bowel necrosis and acute lung injury, and they stimulate the catabolism of muscle protein. In addition, the ability of TNFs to increase the level of collagenase in an arthritic joint and to direct the chemotaxis and migration of leukocytes and lymphocytes may also be responsible for the degradation of cartilage and the proliferation of the synovial tissue in this disease. Therefore, TNFs may serve as mediators of both the acute and chronic stages of immunopathology in rheumatoid arthritis. TNFs may also be responsible for some disorders of blood clotting through altering endothelial cell function. Moreover, excessive TNF production has been demonstrated in patients with AIDS and may be responsible for some of the fever, acute phase response and cachexia seen with this disease and with leukemias.

In these and other circumstances in which TNF has a harmful effect, there is clearly a clinical use for an inhibitor of TNF action. Systemically administered, TNF inhibitors would be useful therapeutics against septic shock and cachexia. Locally applied, such TNF inhibitors would serve to prevent tissue destruction in an inflamed joint and other sites of inflammation. Indeed, such TNF inhibitors could be even more effective when administered in conjunction with interleukin-I (IL-1) inhibitors.

One possibility for therapeutic intervention against the action of TNF is at the level of the target cell's response to the protein. TNF appears to act on

cells through a classical receptor-mediated pathway. Thus, any molecule which interferes with the ability of TNF to bind to its receptors either by blocking the receptor or by blocking the TNF would regulate TNF action. For these reasons, proteins and small molecules capable of inhibiting TNF in this manner have been sought by the present inventors.

SUMMARY OF THE INVENTION

As noted above, this invention relates to TNF inhibitors generally, and, more specifically, to a urine-derived TNF inhibitor. Additionally, the present invention relates to biologically-active analogs of this inhibitor.

An object of the present invention is to provide purified forms of TNF inhibitor which are active against TNF alpha. An additional object of the present invention is to provide these inhibitors in purified forms to enable the determination of their amino acid sequence. A further object is to provide the amino acid sequences of certain TNF inhibitors. In addition it is an object of this invention to provide a cellular source of the mRNA coding for TNF inhibitors and a cDNA library containing a cDNA for the inhibitors. Furthermore, it is an object of this invention to provide a genomic clone of DNA coding for the TNF inhibitors, and the coding sequences of that DNA.

The identification of biologically-active analogs of such TNF inhibitors with enhanced or equivalent properties is also one of the objects of the invention.

Additionally, it is an object of this invention to provide a recombinant-DNA system for the production of the TNF inhibitor described herein. A further object of the present invention includes providing purified forms of TNF inhibitor which would be valuable as pharmaceutical preparations exhibiting activity against TNF. Another object of the present invention includes

providing purified combinations of TNF inhibitors and IL-1 inhibitors which are valuable as pharmaceutical preparations exhibiting activity against both IL-1 and TNF.

The inventors of the present invention have isolated at least two TNF inhibitor proteins with TNF-inhibiting properties. A 30kDa protein and a 40kDa protein have been obtained in their purified forms. The amino acid sequence of the 30kDa TNF inhibitor protein has been obtained. The amino acid sequence data of the 40kDa TNF inhibitor protein has also been obtained. Both the 30kDa TNF inhibitor and the 40kDa TNF inhibitor are novel, previously undescribed proteins.

A human genomic DNA clone which contains the gene for the 30kDa protein has been obtained. A cell source of this protein has been identified and a cDNA clone has been obtained and the nucleic acid sequence of the gene for the protein determined. In addition, the gene clone has been placed in a vector which has been found to express the protein in host cells. Also a process has been developed for purifying the protein in an active form.

A cell source has been identified which produces the 40kDa protein and a cDNA clone has been obtained and the nucleic acid sequence determined of the gene for the 40kDa protein. The full cDNA clones encoding for both the 30kDa TNF inhibitor precursor and the 40kDa TNF inhibitor precursor have been expressed in mammalian cells to yield an increase in TNF binding sites on the cell surface.

A gene coding for the mature form of the 30kDa protein has been expressed in E. Coli. Three seperate genes coding for all or portions of the mature 40kDa protein have also been expressed in E. Coli. The three 40kDa Inhibitor proteins expressed-- mature 40kDa TNF inhibitor, 40kDa TNF inhibitor Δ51 and 40kDa TNF inhibitor Δ53--each exhibit TNF inhibiting activity.

Mature 40kDa TNF inhibitor, as isolated from medium conditioned by human U937 cells, and 40kDa TNF inhibitor Δ51 and 40kDa TNF inhibitor Δ53, are collectively referred to as 40kDa TNF inhibitor.

The 30kDa TNF inhibitor has shown activity in inhibiting TNF alpha. The 40kDa TNF inhibitor has shown inhibitory action against both TNF alpha and TNF beta.

It will now be possible to perform the large scale production of these TNF inhibitors through recombinant DNA technology. These inhibitors should be suitable for use in pharmaceutical formulations useful in treating pathophysiological conditions mediated by TNF.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 describes a cytotoxicity assay for TNF in the absence (-.-.) and in the presence (-x-x-) of TNF inhibitor (30kDa). Various concentrations of TNF were incubated with and without TNF inhibitor, and the cytotoxicity assay was performed as described in Example 1.

Figure 2 describes a native gel shift assay in which "a" depicts TNF alone, and "b" depicts TNF + TNF inhibitor (30kDa).

Figure 3 describes Con A-Peroxidase staining of TNF inhibitor (30kDa). About 200 ng of each protein were run on 14% SDS-PAGE, and transferred to nitrocellulose filter.
Glycoproteins were identified using Con A-peroxidase staining. In this figure, "a" depicts a molecular weight marker, "b" depicts Ovalbumin, "c" depicts bovine serum albumin, and "d" depicts TNF inhibitor.

Figure 4 describes chemical deglycoslylation of TNF inhibitor (30kDa). About 200 ng of TNF inhibitor were chemically deglycosylated (lane C) as described in Example 1.

Figure 5 describes N-glycanase treatment of TNF inhibitor (30kDa). Purified TNF inhibitor was iodinated

by Bolton-Hunter reagent, and denatured-iodinated TNF inhibitor was treated with N-glycanase for 6 hours at 37°C. In this figure, "a" depicts native TNF inhibitor, and "b" depicts deglycosylated TNF inhibitor.

Figure 6A describes an OD_{280} profile of the DEAE Sepharose CL-6B chromatography of 20 l urine.

Figure 6B describes an autoradiograph of the corresponding native gel shift assay indicating a peak of TNF inhibitor at fraction 57-63, which is about 80mM NaCl.

Figure 7 describes an OD_{280} profile of the 0.05 M Na Phosphate pH 2.5 elution from the TNF affinity column.

Figure 8A and 8C describe chromatographic profiles (OD_{215} and OD_{280}) of the RP8 purification of the TNF inhibitor (30kDa) with the L929 bioassay of fractions from the RP8 column showing a peak of TNF inhibitor at fractions 28-31 which is about 18% acetonitrile and at fractions 35 and 36 which is about 21% acetonitrile.

Figure 8B describes a silver stained 15% SDS-PAGE of the RP8 pool showing a single band at 30kDa.

Figure 9A describes a peptide purification of Lys-C digestion of TNF inhibitor (30kDa).

Figure 9B describes a peptide purification of alkylated (*) Lys-C digests of TNF inhibitor (30kDa).

Figure 10 describes a peptide purification of two alkylated (*) Asp-N digests of TNF inhibitor (30kDa).

Figures 11A and 11B describe peptide purifications of an endopeptidase V8 digest of reduced carboxymethylated TNF inhibitor (30kDa).

Figure 12 describes amino acid sequences present in TNF inhibitor (30kDa). Blanks in the sequence indicate the residue has not been unambiguously identified by protein sequencing. C* indicates the identification of carboxymethylcysteine by the presence of ^{3}H in the residue.

Figure 13 describes the DNA sequence of a genomic clone encoding at least a portion of a TNF inhibitor

(30kDa).

Figure 14 describes at least 70% of the mature amino acid sequence of a preferred TNF inhibitor (30kDa).

Figure 15 describes detection of TNF inhibitor in U937 supernatant by the gel shift assay.

Figure 16 describes detection of TNF inhibitor in hplc fractions from U937 supernatant.

Figure 17 describes the Northern blot according to Example 4.

Figure 18 describes the deglycosylated TNF inhibitor (30kDa) binding to TNF. Glycosylated and deglycosylated TNF inhibitor were incubated with TNF affigel, and flow through materials and eluates of the gel were analysed on SDS-PAGE. In this figure, (11) indicates flow through of TNF-INH, reduced and oxidized, (21) indicates flow through of deglycosylated TNF-INH, reduced and oxidized, (51) indicates flow through of native TNF-INH, (12) indicates eluate of TNF-INH, reduced and oxidized, (22) indicates eluate of deglycoslylated TNF-INH, reduced and oxidized, and (52) indicates eluate of native TNF-INH.

D17 Cont'd

Figure 19 describes the complete amino acid sequence of the 30kDa TNF inhibitor.

Figure 20 describes the cDNA sequence encoding the amino acid sequence shown in Figure 19.

Figure 21 describes the entire cDNA sequence for the precursor of the 30kDa TNF inhibitor.

Figure 22 describes the DNA sequence near the start of the TNF inhibitor (30kDa) gene in plasmid pTNFIX-1.

Figure 23 describes the plasmid pCMVXV beta TNFBP stop A.

Figure 24 describes the plasmid pSVXVTNFBP stop A.

Figure 25 describes a chromatographic profile OD_{215} of the RP8 column of the 30kDa TNF inhibitor from E. Coli. The L929 bioassay results are also shown (-x-x).

Figure 26 describes a silver stained 14% SDS-PAGE

of the RP8 Fractions in Figure 25.

Figure 27 describes a chromatographic profile (OD_{215}) of the RP8 purification of the TNF inhibitors from U937 cells. The L929 bioassey results are also shown with a bar graph. Two distinctive TNF inhibitor peaks are seen.

Figure 28 describes a silver stained 14% SDS-PAGE of the RP8 fractions. Fraction number 30 contains the 30kDa TNF inhibitor and fraction number 35 contains the 40kDa TNF inhibitor.

Figure 29 describes a chromatographic profile (OD_{215}) of the purification of urinary 40kDa TNF inhibitor. The second TNF inhibitory peak from several RP8 chromatographies were combined and reanalyzed on an RP8 column. TNF-inhibitory activity is shown with a bar graph. The difference between the OD_{215} peak and the activity peak reflects the dead volume between the detector and the fraction collector.

Figure 30 describes a silver stained 14% SDS-PAGE of the RP8 fractions of urine. Fraction number 32 contains the 40kDa TNF inhibitor.

Figure 31 describes the amino terminal sequences of U937 derived inhibitors (30kDa and 40kDa), and urine-derived 40kDa TNF inhibitor.

Figure 32 describes a peptide purification of endopeptidase V8 digested 40kDa TNF inhibitor.

Figure 33 describes a peptide purification endopeptidase Arg-C digested 40kDa TNF inhibitor.

Figure 34 describes a peptide purification of trypsin digested Arg-C16 peptide.

Figure 35 describes a peptide purification of chymotrypsin digested Arg-C10 peptide.

Figure 36 describes a primary structure of the 40kDa TNF inhibitor.

Figure 37 describes a portion of the 40kDa TNF inhibitor cDNA sequence along with the predicted amino acid translation product.

Figure 38 describes the complete amino acid sequence of the 40kDa TNF inhibitor.

Figure 39 describes the entire cDNA sequence for the precursor of the 40kDa TNF inhibitor, along with its deduced translation product.

Figure 40 describes a cytotoxicity assay for TNF beta (lymphotoxin) in the presence (o-o) of 40kDa TNF inhibitor, in the presense (●-●) of 30kDa TNF inhibitor and without any inhibitor (x-x).

Figure 41 describes the expression of the 30kDa TNF inhibitor cDNA sequence shown in Figure 21 in COS7 cells. COS cells were transfected with plasmids using the lipofectin procedure of Feigner et al. (Proc. Natl. Acad. Sci. (USA) 84, 7413-1987). 3.4×10^5 cells were incubated with the indicated amounts of [^{125}I] TNFa at a specific activity of 5.6×10^4 cpm/ng and the amount bound to the cells determined. Open symbols are the total cpm associated with cells after a 4 hour incubation at 4°C. Closed symbols represent bound [^{125}I] TNFa in the presense of 180 fold excess of cold unlabeled TNFa.

Figure 42 describes the expression of the 40kDa TNF inhibitor cDNA sequence shown in Figure 39 in COS7. Assay conditions were as described in Figure 41. The darkened symbols represent the bound [^{125}I] TNFa in the presense of 180 fold excess of cold unlabeled TNFa.

Figure 43 describes the cytotoxicity assay of an HPLC RPC-8 fraction of the human monocytes which were treated with PMA and PHA for 24 hours.

Figure 44 describes the RPC-8 chromatographic pattern of 40kDa TNF inhibitor Δ51, SDS-polyacrylamide gel analysis of the fractions (B), and the cytotoxicity assay on L929 cells (C).

Figure 45 describes the RPC-8 chromatographic pattern of 40kDa TNF inhibitor Δ53 (A), SDS-polyacrylamide gel analysis of the fractions (B), and the cytotoxicity assay on L929 cells (C).

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Reference will now be made in detail to the presently preferred embodiments of the invention, which, together with the following examples, serve to explain the principals of the invention.

1. Inhibitor isolated from urine.

As noted above, the present invention relates to TNF inhibitors which have been isolated in a purified form. In one embodiment of this invention, the TNF inhibitors are preferably derived from urine. In addition, the invention encompasses substantially purified TNF inhibitors of any origin which are biologically equivalent to the inhibitor isolated from urine. Throughout this specification, any reference to a TNF inhibitor or simply an inhibitor should be construed to refer to each of the inhibitors identified and described herein.

By "biologically equivalent" as used throughout the specification and claims, we mean compositions of the present invention which are capable of preventing TNF action in a similar fashion, but not necessarily to the same degree as the native TNF inhibitor isolated from urine. By "substantially homologous" as used throughout the ensuing specification and claims, is meant a degree of homology to the native TNF inhibitor isolated from urine in excess of that displayed by any previously reported TNF inhibitor. Preferably, the degree of homology is in excess of 70%, most preferably in excess of 80%, and even more preferably in excess of 90%, 95% or 99%. A particularly preferred group of TNF inhibitors are in excess of 95% homologous with the native inhibitor. The percentage of homology as described herein is calculated as the percentage of amino acid residues found in the smaller of the two

sequences which align with identical amino acid residues in the sequence being compared when four gaps in a length of 100 amino acids may be introduced to assist in that alignment as set forth by Dayhoff, in Atlas of Protein Sequence and Structure Vol. 5, p. 124 (1972), National Biochemical Research Foundation, Washington, D.C., specifically incorporated herein by reference. Also included as substantially homologous are those TNF inhibitors which may be isolated by virtue of cross-reactivity with antibodies to the described inhibitor or whose genes may be isolated through hybridization with the gene or with segments of the described inhibitor.

The preferred TNF inhibitors of the present invention have been derived from urine and, for the first time, have been isolated in a purified form. For the purposes of the present application, "pure form" or "purified form," when used to refer to the TNF inhibitors disclosed herein, shall mean a preparation which is substantially free of other proteins which are not TNF inhibitor proteins. Preferably, the TNF inhibitors of the present invention are at least 50% percent pure, preferably 75% pure and more preferably 80%, 95% or 99% pure. In one embodiment of the present invention, the TNF inhibitor protein preparation is sufficiently pure to enable one of ordinary skill in the art to determine its amino acid sequence without first performing further purification steps.

At least two TNF inhibitors have been isolated by the methods set forth in the examples. The two inhibitors are approximately 30kDa and approximately 40kDa molecules on SDS-PAGE. The 30kDa inhibitor elutes from a DEAE CL6B column at about 80 millimolar NaCl in Tris buffer, pH 7.5. The amino acid sequence of the 30kDa inhibitor is set forth in Figure 19, and the amino acid sequence of the 40kDa inhibitor is set forth in Figure 38. The 30kDa TNF inhibitor has been shown to inhibit the activity of TNF alpha, and has little effect

on the activity of TNF beta. The 40kDa TNF inhibitor has been shown to exhibit a significant inhibiting effect against both TNF alpha and TNF beta (lymphotoxin).

2. Inhibitor isolated from U937 conditioned medium

In an alternate embodiment of the present invention, TNF inhibitors are isolated from a medium conditioned by human U937 cells. Two TNF inhibitor proteins have been identified and isolated from this conditioned medium. The two TNF inhibitors are 30kDa and 40kDa proteins that are substantially homologous to the 30kDa and 40kDa TNF inhibitors isolated from urine, and are biologically equivalent to such proteins.

3. Structure of 30kDa TNF inhibitor

The 30kDa TNF inhibitor isolated from urine is a glycoprotein, containing at least one carbohydrate moiety. In one embodiment of this invention, the natural 30kDa TNF inhibitor is deglycosylated. The deglycosylated TNF inhibitor, which retains its ability to bind to TNF, is within the scope of the present invention. Fully and partially deglycosylated 30kDa TNF inhibitor is encompassed by this invention. The deglycosylated 30kDa TNF inhibitor isolated from urine is about an 18kDa protein.

The gene sequence identified that encodes the 30kDa protein does not contain a termination codon as would be anticipated for the amino acid sequence of the 18kDa protein. The inventors theorize, but are in no way to be limited by this theory, that the proteins produced in vivo contain additional amino acid sequences. According to this theory, the protein encoded is a TNF receptor molecule. The inhibitor protein encoded by the cDNA has a hydrophobic sequence that would be compatible with the cell membrane spanning region and a TNF binding portion that would extend extracellularly from the cell

membrane. In accord with this hypothesis and as described in Example 19, the cDNA has been expressed in COS cells and leads to an increase in the number of TNF binding sites on the cell. The TNF inhibitors of the present invention, therefore, are the receptor fragments or portions of the receptor molecule. Such binding fragments have been identified with respect to other binding/inhibitory molecules (e.g., IL-2 inhibitor), and are referred to as soluble receptors.

This theory is consistent with the lack of a termination codon in the nucleotide sequence that would correspond to the terminus of the protein as anticipated by the known sequence of the isolated TNF inhibitor factor. It is also consistent with the fact that the nucleotide sequence beyond where the termination codon should be found, encodes a series of hydrophobic amino acids.

The present invention, therefore, encompasses not just the portion of the TNF inhibitors identified and described, but all proteins containing any portion of the amino acid sequence encoded by the cDNA sequences identified and described herein.

4. Structure of 40kDa TNF inhibitor.

The 40kDa TNF inhibitor isolated from medium conditioned by human U937 cells and identified in urine is a glycoprotein, containing at least one carbohydrate moiety. In one embodiment of this invention, the natural 40kDa TNF inhibitor is deglycosylated. The deglycosylated 40kDa TNF inhibitor, which retains its ability to bind to both TNF alpha and TNF beta (lymphotoxin) is within the scope of the present invention. Fully and partially deglycosylated 40kDa TNF inhibitor is encompassed by this invention. The inventors theorize that the 40kDa TNF inhibitor may also be a soluble receptor. The gene sequence identified that encodes the 40kDa protein does not contain a

termination codon as would be anticipated for the amino acid sequence of the deglycosylated 40kDa TNF inhibitor. As described in Example 20, the cDNA has been expressed in COS cells and leads to an increase in the number of TNF binding sites on the cell.

The present invention encompasses the gene encoding the mature 40kDa protein isolated from medium conditioned by human U937 cells and identified in urine, and larger and smaller portions of such gene to the extent that the TNF inhibiting activity of the encoded protein is not effected. As can be seen by reference to Figure 38, the mature 40kDa TNF inhibitor has a proline rich area near the anticipated c-terminus of the protein. 40kDa TNF inhibitors in which all or portions of the proline rich regime are not included in the protein are active as TNF inhibitors, and are within the scope of the present invention. Two such shortened proteins are described in Examples 17 and 22 below, and are referred to as 40kDa TNF inhibitor Δ51 and 40kDa TNF inhibitor Δ53. All portions of this application which refer generally to 40kDa TNF inhibitor shall encompass the mature 40kDa protein isolated from medium conditioned by human U937 cells and identified in urine, as well as 40kDa TNF inhibitor Δ51 and 40kDa TNF inhibitor Δ53.

It is generally believed that at least one TNF receptor is capable of binding both TNF alpha and TNF beta, while some TNF receptors are capable of only binding TNF alpha. This is consistent with the findings in the present invention wherein two TNF inhibitors have been identified which are both proposed to be active fragments of TNF receptor sites, and one is active against only TNF alpha and the other is active against both TNF alpha and TNF beta.

5. Recombinant inhibitor.

(a) General

A recombinant DNA method for the manufacture of a

TNF inhibitor is now disclosed. In one embodiment of the invention, the active site functions in a manner biologically equivalent to that of the TNF inhibitor isolated from urine. A natural or synthetic DNA sequence may be used to direct production of such TNF inhibitors. This method comprises:

(a) preparation of a DNA sequence capable of directing a host cell to produce a protein having TNF inhibitor activities or a precursor thereof;

(b) cloning the DNA sequence into a vector capable of being transferred into and replicated in a host cell, such vector containing operational elements needed to express the DNA sequence or a precursor thereof;

(c) transferring the vector containing the synthetic DNA sequence and operational elements into a host cell capable of expressing the DNA encoding TNF inhibitor or a precursor thereof;

(d) culturing the host cells under the conditions for amplification of the vector and expression of the inhibitor or a precursor thereof;

(e) harvesting the inhibitor or a precursor thereof; and

(f) permitting the inhibitor to assume an active tertiary structure whereby it possesses or can be processed into a protein having TNF inhibitory activity.

In one embodiment of the present invention, the TNF inhibitor is produced by the host cell in the form of a precursor protein. This precursor protein is processed to a protein in one or more steps and allowed to fold correctly to an active TNF inhibitor using methods generally known to those of ordinary skill in the art.

(b) <u>DNA sequences</u>

DNA sequences contemplated for use in this method are discussed in part in Examples 6, 14A, and 17. It is contemplated that these sequences include synthetic and natural DNA sequences and combinations thereof. The

natural sequences further include cDNA or genomic DNA segments.

The means for synthetic creation of polynucleotide sequences encoding a protein identical to that encoded by the cDNA or genomic polynucleotide sequences are generally known to one of ordinary skill in the art, particularly in light of the teachings contained herein. As an example of the current state of the art relating to polynucleotide synthesis, one is directed to Matteucci, M.D., and Caruthers, M.H., in J. Am. Chem. Soc. 103:3185 (1981) and Beaucage, S.L. and Caruthers, M.H. in Tetrahedron Lett. 22:1859 (1981), and to the instructions supplied with an ABI oligonucleotide synthesizer, each of which is specifically incorporated herein by reference.

These synthetic sequences may be identical to the natural sequences described in more detail below or they may contain different nucleotides. In one embodiment, if the synthetic sequences contain nucleotides different from those found in the natural DNA sequences of this invention, it is contemplated that these different sequences will still encode a polypeptide which has the same primary structure as TNF inhibitor isolated from urine. In an alternate embodiment, the synthetic sequence containing different nucleotides will encode a polypeptide which has the same biological activity as the TNF inhibitor described herein.

Additionally, the DNA sequence may be a fragment of a natural sequence, i.e., a fragment of a polynucleotide which occurred in nature and which has been isolated and purified for the first time by the present inventors. In one embodiment, the DNA sequence is a restriction fragment isolated from a cDNA library.

In an alternative embodiment, the DNA sequence is isolated from a human genomic library. An example of such a library useful in this embodiment is set forth by Wyman, et al., (1985) Proc. Nat. Acad. Sci. USA, 82,

2880-2884.

In a preferred version of this embodiment, it is contemplated that the natural DNA sequence will be obtained by a method comprising:

(a) Preparation of a human cDNA library from cells, preferably U937 cells capable of generating a TNF inhibitor, in a vector and a cell capable of amplifying and expressing all or part of that cDNA;

(b) Probing the human DNA library with at least one probe capable of binding to the TNF inhibitor gene or its protein product;

(c) Identifying at least one clone containing the gene coding for the inhibitor by virtue of the ability of the clone to bind at least one probe for the gene or its protein product;

(d) Isolation of the gene or portion of the gene coding for the inhibitor from the clone or clones chosen; and

(e) Linking the gene, or suitable fragments thereof, to operational elements necessary to maintain and express the gene in a host cell.

The natural DNA sequences useful in the foregoing process may also be identified and isolated through a method comprising:

(a) Preparation of a human genomic library, preferably propagated in a recBC,sbc host, preferably CES 200;

(b) Probing the human genomic library with at least one probe capable of binding a TNF inhibitor gene or its protein product;

(c) Identification of at least one clone containing the gene coding for the inhibitor by virtue of the ability of the clone to bind at least one probe for the gene or its protein product;

(d) Isolation of the gene coding for the inhibitor from the clone or clones identified; and

(e) Linking the gene, or suitable fragments

thereof, to operational elements to maintain and express the gene in a host cell.

A third potential method for identifying and isolating natural DNA sequences useful in the foregoing process includes the following steps:

(a) Preparation of mRNA from cells that produce the TNF inhibitor;

(b) Synthesizing cDNA (single- or double-stranded) from this mRNA;

(c) Amplifying the TNF inhibitor-specific DNA sequences present in this mixture of cDNA sequences using the polymerase chain reaction (PCR) procedure with primers such as those shown in Table 5;

(d) Identifying the PCR products that contain sequences present in the other oligonucleotide probes shown in Table 5 using Southern blotting analysis;

(e) Subcloning the DNA fragments so identified into M13 vectors that allow direct sequencing of the DNA sequences;

(f) Using these sequences to isolate a cDNA clone from a cDNA library; and

(g) Linking the gene, or suitable fragments thereof, to operational elements necessary to maintain and express the gene in host cells.

In isolating a DNA sequence suitable for use in the above-method, it is preferred to identify the two restriction sites located within and closest to the end portions of the appropriate gene or sections of the gene that encode the native protein or fragments thereof. The DNA segment containing the appropriate gene or sections of the gene is then removed from the remainder of the genomic material using appropriate restriction endonucleases. After excision, the 3' and 5' ends of the DNA sequence and any intron exon junctions are reconstructed to provide appropriate DNA sequences

capable of coding for the N- and C- termini and the body of the TNF inhibitor protein and capable of fusing the DNA sequence to its operational elements.

As described in Example 17 below, the DNA sequence utilized for the expression of 40kDa TNF inhibitor may be modified by the removal of either 153 or 159 base pairs from the gene that encodes for the mature 40kDa TNF inhibitor isolated from medium conditioned by human U937 cells and identified in urine. The Δ53 gene was prepared to remove the proline regime from the C-terminus of the full gene, and the Δ51 gene was prepared to approximate the C-terminus of the gene encoding for 30kDa TNF inhibitor.

A DNA sequence, isolated according to these methods from a cDNA library and encoding at least a portion of the 30kDa TNF inhibitor described herein, has been deposited at the American Type Culture Collection, Rockville, M.D., under Accession No 40645.

A DNA sequence, isolated according to these methods from a human genomic DNA library and encoding at least a portion of the 30kDa TNF inhibitor described herein, has been deposited at the American Type Culture Collection, Rockville, MD., under Accession No. 40620.

A DNA sequence, isolated according to these methods from a cDNA library and encoding at least a portion of the 40kDa TNF inhibitor described herein, has been deposited at the American Type Culture Collection, Rockville, MD, under Accession No. 68204.

6. Vectors

(a) Microorganisms, especially E. coli

The vectors contemplated for use in the present invention include any vectors into which a DNA sequence as discussed above can be inserted, along with any preferred or required operational elements, and which vector can then be subsequently transferred into a host cell and replicated in such cell. Preferred vectors are

those whose restriction sites have been well documented and which contain the operational elements preferred or required for transcription of the DNA sequence. However, certain embodiments of the present invention are also envisioned which employ currently undiscovered vectors which would contain one or more of the cDNA sequences described herein. In particular, it is preferred that all of these vectors have some or all of the following characteristics: (1) possess a minimal number of host-organism sequences; (2) be stably maintained and propagated in the desired host; (3) be capable of being present in a high copy number in the desired host; (4) possess a regulatable promoter positioned so as to promote transcription of the gene of interest; (5) have at least one marker DNA sequence coding for a selectable trait present on a portion of the plasmid separate from that where the DNA sequence will be inserted; and (6) a DNA sequence capable of terminating transcription.

In variously preferred embodiments, these cloning vectors containing and capable of expressing the DNA sequences of the present invention contain various operational elements. These "operational elements," as discussed herein, include at least one promoter, at least one Shine-Dalgarno sequence and initiator codon, and at least one terminator codon. Preferably, these "operational elements" also include at least one operator, at least on leader sequence for proteins to be exported from intracellular space, at least one gene for a regulator protein, and any other DNA sequences necessary or preferred for appropriate transcription and subsequent translation of the vector DNA.

Certain of these operational elements may be present in each of the preferred vectors of the present invention. It is contemplated that any additional operational elements which may be required may be added to these vector using methods known to those of ordinary

skill in the art, particularly in light of the teachings herein.

In practice, it is possible to construct each of these vectors in a way that allows them to be easily isolated, assembled and interchanged. This facilitates assembly of numerous functional genes from combinations of these elements and the coding region of the DNA sequences. Further, many of these elements will be applicable in more than one host. It is additionally contemplated that the vectors, in certain preferred embodiments, will contain DNA sequences capable of functioning as regulators ("operators"), and other DNA sequences capable of coding for regulator proteins.

(i) Regulators

These regulators, in one embodiment, will serve to prevent expression of the DNA sequence in the presence of certain environmental conditions and, in the presence of other environmental conditions, will allow transcription and subsequent expression of the protein coded for by the DNA sequence. In particular, it is preferred that regulatory segments be inserted into the vector such that expression of the DNA sequence will not occur, or will occur to a greatly reduced extent, in the absence of, for example, isopropylthio-beta-D-galactoside. In this situation, the transformed microorganisms containing the DNA sequence may be grown to a desired density prior to initiation of the expression of TNF inhibitor. In this embodiment, expression of the desired protein is induced by addition of a substance to the microbial environment capable of causing expression of the DNA sequence after the desired density has been achieved.

(ii) Promoters

The expression vectors must contain promoters which can be used by the host organism for expression of its own proteins. While the lactose promoter system is commonly used, other microbial promoters have been

isolated and characterized, enabling one skilled in the art to use them for expression of the recombinant TNF inhibitor.

(iii) Transcription Terminator

The transcription terminators contemplated herein serve to stabilize the vector. In particular, those sequences as described by Rosenberg, M. and Court, D., in Ann. Rev. Genet. 13:319-353 (1979), specifically incorporated herein by reference, are contemplated for use in the present invention.

(iv) Non-Translated Sequence

It is noted that, in the preferred embodiment, it may also be desirable to reconstruct the 3' or 5' end of the coding region to allow incorporation of 3' or 5' non-translated sequences into the gene transcript. Included among these non-translated sequences are those which stabilize the mRNA as they are identified by Schmeissner, U., McKenney, K., Rosenberg, M and Court, D. in J. Mol. Biol. 176:39-53 (1984), specifically incorporated herein by reference.

(v) Ribosome Binding Sites

The microbial expression of foreign proteins requires certain operational elements which include, but are not limited to, ribosome binding sites. A ribosome binding site is a sequence which a ribosome recognizes and binds to in the initiation of protein synthesis as set forth in Gold, L., et al., Ann. Rev. Microbio. 35:557-580; or Marquis, D.M., et al., Gene 42:175-183 (1986), both of which are specifically incorporated herein by reference. A preferred ribosome binding site is GAGGCGCAAAAA(ATG).

(vi) Leader Sequence and Translational Coupler

Additionally, it is preferred that DNA coding for an appropriate secretory leader (signal) sequence be present at the 5' end of the DNA sequence as set forth by Watson, M.E. in Nucleic Acids Res. 12:5145-5163, specifically incorporated herein by reference, if the

protein is to be excreted from the cytoplasm. The DNA for the leader sequence must be in a position which allows the production of a fusion protein in which the leader sequence is immediately adjacent to and covalently joined to the inhibitor, i.e., there must be no transcription or translation termination signals between the two DNA coding sequences. The presence of the leader sequence is desired in part for one or more of the following reasons. First, the presence of the leader sequence may facilitate host processing of the TNF inhibitor. In particular, the leader sequence may direct cleavage of the initial translation product by a leader peptidase to remove the leader sequence and leave a polypeptide with the amino acid sequence which has potential protein activity. Second, the presence of the leader sequence may facilitate purification of the TNF inhibitor, through directing the protein out of the cell cytoplasm. In some species of host microorganisms, the presence of an appropriate leader sequence will allow transport of the completed protein into the periplasmic space, as in the case of some E. coli. In the case of ceratin E. coli, Saccharomyces and strains of Bacillus and Pseudomonas, the appropriate leader sequence will allow transport of the protein through the cell membrane and into the extracellular medium. In this situation, the protein may be purified from extracellular protein. Thirdly, in the case of some of the proteins prepared by the present invention, the presence of the leader sequence may be necessary to locate the completed protein in an environment where it may fold to assume its active structure, which structure possesses the appropriate protein activity.

In one preferred embodiment of the present invention, an additional DNA sequence is located immediately preceding the DNA sequence which codes for the TNF inhibitor. The additional DNA sequence is capable of functioning as a translational coupler, i.e.,

it is a DNA sequence that encodes an RNA which serves to position ribosomes immediately adjacent to the ribosome binding site of the inhibitor RNA with which it is contiguous. In one embodiment of the present invention, the translational coupler may be derived using the DNA sequence TAACGAGGCGCAAAAAATGAAAAAGACAGCTATCGCGATCTTGGAGGATGATTAAATG and methods currently known to those of ordinary skill in the art related to translational couplers.

(vii) Translation Terminator

The translation terminators contemplated herein serve to stop the translation of mRNA. They may be either natural, as described by Kohli, J., Mol. Gen. Genet. 182:430-439; or synthesized, as described by Pettersson, R.F. Gene 24:15-27 (1983), both of which references are specifically incorporated herein by reference.

(viii) Selectable Marker

Additionally, it is preferred that the cloning vector contain a selectable marker, such as a drug resistance marker or other marker which causes expression of a selectable trait by the host microorganism. In one embodiment of the present invention, the gene for ampicillin resistance is included in the vector while, in other plasmids, the gene for tetracycline resistance or the gene for chloramphenicol resistance is included.

Such a drug resistance or other selectable marker is intended in part to facilitate in the selection of transformants. Additionally, the presence of such a selectable marker in the cloning vector may be of use in keeping contaminating microorganisms from multiplying in the culture medium. In this embodiment, a pure culture of the transformed host microorganisms would be obtained by culturing the microorganisms under conditions which require the induced phenotype for survival.

The operational elements as discussed herein are routinely selected by those or ordinary skill in the art

in light of prior literature and the teachings contained herein. General examples of these operational elements are set forth in B. Lewin, Genes, Wiley & Sons, New York (1983), which is specifically incorporated herein by reference. Various examples of suitable operational elements may be found on the vectors discussed above and may be elucidated through review of the publications discussing the basic characteristics of the aforementioned vectors.

Upon synthesis and isolation of all necessary and desired component parts of the above-discussed vector, the vector is assembled by methods generally known to those of ordinary skill in the art. Assembly of such vectors is believed to be within the duties and tasks performed by those with ordinary skill in the art and, as such, is capable of being performed without undue experimentation. For example, similar DNA sequences have been ligated into appropriate cloning vectors, as set forth by Maniatis et al. in Molecular Cloning, Cold Spring Harbor Laboratories (1984), which is specifically incorporated herein by reference.

In construction of the cloning vectors of the present invention, it should additionally be noted that multiple copies of the DNA sequence and its attendant operational elements may be inserted into each vector. In such an embodiment, the host organism would produce greater amounts per vector of the desired TNF inhibitor. The number of multiple copies of the DNA sequence which may be inserted into the vector is limited only by the ability of the resultant vector, due to its size, to be transferred into and replicated and transcribed in an appropriate host cell.

(b) Other Microorganisms

Vectors suitable for use in microorganisms other than E. coli are also contemplated for this invention. Such vectors are described in Table 1. In addition, certain preferred vectors are discussed below.

TABLE 1

HOSTS	REGULATED PROMOTERS	INDUCER	TRANSCRIPTION TERMINATOR	MRNA STABILIZATION	TRANSCRIPTIONAL START SITE & LEADER PEPTIDE	MARKER	RS BINDING SITE
E.coli	Lac[1], Tac[2] Lambda pL Trp[3]	IPTG increased temperature IAA addition or tryptophan depletion	rrnB[6] rrnC[7]	ompA[8] lambda int[9] trp[10]	bla[11] ompA[12] phoS	ampicillin[14] tetracycline[14,15] chloramphenical[16]	
Bacillus	*alpha amylase[17] *subtilisin[18] *p-43[19] spac-1[26]	IPTG	E. coli rrn rrn BT.T[20]		B.amy neutral protease[21] B.amy alpha-amylase[22] B.subt. subtilisin[23]	Kan^r 24 Cam^r 25	B. amy neural protease B.amy alpha-amylase[22]
Pseudo-monas	Trp[27](E.coli) Lac(E.coli) Tac(E.coli)	IAA addition, or tryptophan depletion IPTG			phospholipase C28 exotoxin A[28]	sulfonamide[30] streptomycin[30]	Trp (E.coli)
Yeast	Gal 1[31], 10[32] Adh I[33], II[34] Pho 5	Glucose depletion and galactose Glucose depletion Phosphate depletion	Cyc1 Una Alpha factor Sac 2		Invertase[36] Acid phosphatase[36] Alpha Factor	Ura 3[37] Leu 2[38] His 3 Tap 1	

*non-regulated

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(i) Pseudomonas Vectors

Several vector plasmids which autonomously replicate in a broad range of Gram negative bacteria are preferred for use as cloning vehicles in hosts of the genus Pseudomonas. Certain of these are described by Tait, R.C., Close, T.J., Lundquist, R.C., Hagiya, M., Rodriguez, R.L., and Kado, C.I. in Biotechnology, May, 1983, pp. 269-275; Panopoulos, N.J. in Genetic Engineering in the Plant Sciences, Praeger Publishers, New York, New York, pp. 163-185 (1981); and Sakagucki, K. in Current Topics in Microbiology and Immunology 96:31-45 (1982), each of which is specifically incorporated herein by reference.

One particularly preferred construction would employ the plasmid RSF1010 and derivatives thereof as described by Bagdasarian, M., Bagdasarian, M.M., Coleman, S., and Timmis, K.N. in Plasmids of Medical, Environmental and Commercial Importance, Timmis, K.N. and Puhler, A. eds., Elsevier/North Holland Biomedical Press (1979), specifically incorporated herein by reference. The advantages of RSF1010 are that it is a relatively small, high copy number plasmid which is readily transformed into and stably maintained in both E. coli and Pseudomonas species. In this system, it would be preferred to use the Tac expression system as described for Escherichia, since it appears that the E. coli trp promoter is readily recognized by Pseudomonas RNA polymerase as set forth by Sakagucki, K. in Current Topics in Microbiology and Immunology 96:31-45 (1982) and Gray, G.L., McKeown, K.A., Jones A.J.S., Seeburg, P.H., and Heyneker, H.L. in Biotechnology, Feb. 1984, pp. 161-165, both of which are specifically incorporated herein by reference. Transcriptional activity may be further maximized by requiring the exchange of the promoter with, e.g., an E. coli or P. aeruginosa trp promoter. Additionally, the lacI gene of E. coli would also be

included in the plasmid to effect regulation.

Translation may be coupled to translation initiation for any of the Pseudomonas proteins, as well as to initiation sites for any of the highly expressed proteins of the type chosen to cause intracellular expression of the inhibitor.

In those cases where restriction minus strains of a host Pseudomonas species are not available, transformation efficiency with plasmid constructs isolated from E. coli are poor. Therefore, passage of the Pseudomonas cloning vector through an r- m+ strain of another species prior to transformation of the desired host, as set forth in Bagdasarian, M., et al., Plasmids of Medical, Environmental and Commercial Importance, pp. 411-422, Timmis and Puhler eds., Elsevier/North Holland Biomedical Press (1979), specifically incorporated herein by reference, is desired.

(ii) Bacillus Vectors

Furthermore, a preferred expression system in hosts of the genus Bacillus involves using plasmid pUB110 as the cloning vehicle. As in other host vector systems, it is possible in Bacillus to express the TNF inhibitor of the present invention as either an intracellular or a secreted protein. The present embodiments include both systems. Shuttle vectors that replicate in both Bacillus and E. coli are available for constructing and testing various genes as described by Dubnau, D., Gryczan, T., Contente, S., and Shivakumar, A.G. in Genetic Engineering, Vol. 2, Setlow and Hollander eds., Plenum Press, New York, New York, pp. 115-131 (1980), specifically incorporated herein by reference. For the expression and secretion of the TNF inhibitor from B. subtilis, the signal sequence of alpha-amylase is preferably coupled to the coding region for the protein. For synthesis of intracellular inhibitor, the portable DNA sequence will be translationally coupled to the ribosome binding site of the alpha-amylase leader

sequence.

Transcription of either of these constructs is preferably directed by the alpha-amylase promoter or a derivative thereof. This derivative contains the RNA polymerase recognition sequence of the native alpha-amylase promoter but incorporates the lac operator region as well. Similar hybrid promoters constructed from the penicillinase gene promoter and the lac operator have been shown to function in Bacillus hosts in a regulatable fashion as set forth by Yansura, D.G. and Henner in Genetics and Biotechnology of Bacilii, Ganesan, A.T. and Hoch, J.A., eds., Academic Press, pp. 249-263 (1984), specifically incorporated by reference. The lacI gene of E. coli would also be included in the plasmid to effect regulation.

(iii) Clostridium Vectors

One preferred construction for expression in Clostridium is in plasmid pJU12, described by Squires, C.H. et al., in J. Bacteriol. 159:465-471 (1984) and specifically incorporated herein by reference, transformed into C. perfringens by the method of Heefner, D.L. et al., as described in J. Bacteriol. 159:460-464 (1984), specifically incorporated herein by reference. Transcription is directed by the promoter of the tetracycline resistance gene. Translation is coupled to the Shine-Dalgarno sequences of this same tet^r gene in a manner strictly analogous to the procedures outlined above for vectors suitable for use in other hosts.

(iv) Yeast Vectors

Maintenance of foreign DNA introduced into yeast can be effected in several ways as described by Botstein, D. and Davis, R.W., in The Molecular Biology of the Yeast Saccharomyces, Cold Spring Harbor Laboratory, Strathern, Jones and Broach, eds., pp. 607-636 (1982), specifically incorporated herein by reference. One preferred expression system for use with host organisms of the genus Saccharomyces harbors the TNF inhibitor gene on the

2 micron plasmid. The advantages of the 2 micron circle include relatively high copy number and stability when introduced into cir' strains. These vectors preferably incorporate the replication origin and at least one antibiotic resistance marker from pBR322 to allow replication and selection in E. coli. In addition, the plasmid will preferably have the two micron sequence and the yeast LEU2 gene to serve the same purposes in LEU2 defective mutants of yeast.

If it is contemplated that the recombinant TNF inhibitors will ultimately be expressed in yeast, it is preferred that the cloning vector first be transferred into Escherichia coli, where the vector would be allowed to replicate and from which the vector would be obtained and purified after amplification. The vector would then be transferred into the yeast for ultimate expression of the TNF inhibitor.

(c) Mammalian Cells

The cDNA for the TNF inhibitor will serve as the gene for expression of the inhibitor in mammalian cells. It should have a sequence that will be efficient at binding ribosomes such as that described by Kozak, in Nucleic Acids Research 15:8125-8132 (1987), specifically incorporated herein by reference, and should have coding capacity for a leader sequence (see section 3(a)(vi)) to direct the mature protein out of the cell in a processed form. The DNA restriction fragment carrying the complete cDNA sequence can be inserted into an expression vector which has a transcriptional promoter and a transcriptional enhancer as described by Guarente, L. in Cell 52:303-305 (1988) and Kadonaga, J.T. et al., in Cell 51:1079-1090 (1987), both of which are specifically incorporated herein by reference. The promoter may be regulatable as in the plasmid pMSG (Pharmacia Cat. No. 27450601) if constitutive expression of the inhibitor is harmful to cell growth. The vector should have a complete polyadenylation signal as described by Ausubel,

F.M. et al. in Current Protocols in Molecular Biology, Wiley (1987), specifically incorporated herein by reference, so that the mRNA transcribed from this vector is processed properly. Finally, the vector will have the replication origin and at least one antibiotic resistance marker from pBR322 to allow replication and selection in E. coli.

In order to select a stable cell line that produces the TNF inhibitor, the expression vector can carry the gene for a selectable marker such as a drug resistance marker or carry a complementary gene for a deficient cell line, such as a dihydrofolate reductase (dhfr) gene for transforming a dhfr$^-$ cell line as described by Ausubel et al., supra. Alternatively, a separate plasmid carrying the selectable marker can be cotransformed along with the expression vector.

7. Host Cells/Transformation

The vector thus obtained is transferred into an appropriate host cell. These host cells may be microorganisms or mammalian cells.

(α) (c) Microorganisms

It is believed that any microorganism having the ability to take up exogenous DNA and express those genes and attendant operational elements may be chosen. After a host organism has been chosen, the vector is transferred into the host organism using methods generally known to those of ordinary skill in the art. Examples of such methods may be found in Advanced Bacterial Genetics by R.W. Davis et al., Cold Spring Harbor Press, Cold Spring Harbor, New York, (1980), which is specifically incorporated herein by reference. It is preferred, in one embodiment, that the transformation occur at low temperatures, as temperature regulation is contemplated as a means of regulating gene expression through the use of operational elements as set forth above. In another embodiment, if osmolar regulators have

been inserted into the vector, regulation of the salt concentrations during the transformation would be required to insure appropriate control of the foreign genes.

It is preferred that the host microorganism be a facultative anaerobe or an aerobe. Particular hosts which may be preferable for use in this method include yeasts and bacteria. Specific yeasts include those of the genus Saccharomyces, and especially Saccharomyces cerevisiae. Specific bacteria include those of the genera Bacillus, Escherichia, and Pseudomonas, especially Bacillus subtilis and Escherichia coli. Additional host cells are listed in Table I, supra.

(b) ~~(a)~~ Mammalian Cells

The vector can be introduced into mammalian cells in culture by several techniques such as calcium phosphate: DNA coprecipitation, electroporation, or protoplast fusion. The preferred method is coprecipitation with calcium phosphate as described by Ausubel et al., supra.

Many stable cell types exist that are transformable and capable of transcribing and translating the cDNA sequence, processing the precursor TNF inhibitor and secreting the mature protein. However, cell types may be variable with regard to glycosylation of secreted proteins and post-translational modification of amino acid residues, if any. Thus, the ideal cell types are those that produce a recombinant TNF inhibitor identical to the natural molecule.

8. Culturing Engineered Cells

The host cells are cultured under conditions appropriate for the expression of the TNF inhibitor. These conditions are generally specific for the host cell, and are readily determined by one of ordinary skill in the art in light of the published literature regarding the growth conditions for such cells and the teachings contained herein. For example, Bergey's Manual of

Determinative Bacteriology, 8th Ed., Williams & Wilkins Company, Baltimore, Maryland, which is specifically incorporated herein by reference, contains information on conditions for culturing bacteria. Similar information on culturing yeast and mammalian cells may be obtained from Pollack, R. Mammalian Cell Culture, Cold Spring Harbor Laboratories (1975), specifically incorporated herein by reference.

Any conditions necessary for the regulation of the expression of the DNA sequence, dependent upon any operational elements inserted into or present in the vector, would be in effect at the transformation and culturing stages. In one embodiment, cells are grown to a high density in the presence of appropriate regulatory conditions which inhibit the expression of the DNA sequence. When optimal cell density is approached, the environmental conditions are altered to those appropriate for expression of the DNA sequence. It is thus contemplated that the production of the TNF inhibitor will occur in a time span subsequent to the growth of the host cells to near optimal density, and that the resultant TNF inhibitor will be harvested at some time after the regulatory conditions necessary for its expression were induced.

9. Purification

(a) TNF inhibitor Produced From Microorganisms.

In a preferred embodiment of the present invention, the recombinant TNF inhibitor is purified subsequent to harvesting and prior to assumption of its active structure. This embodiment is preferred as the inventors believe that recovery of a high yield of re-folded protein is facilitated if the protein is first purified. However, in one preferred, alternate embodiment, the TNF inhibitor may be allowed to refold to assume its active structure prior to purification. In yet another preferred, alternate embodiment, the TNF inhibitor is

present in its re-folded, active state upon recovery from the culturing medium.

In certain circumstances, the TNF inhibitor will assume its proper, active structure upon expression in the host microorganism and transport of the protein through the cell wall or membrane or into the periplasmic space. This will generally occur if DNA coding for an appropriate leader sequence has been linked to the DNA coding for the recombinant protein. If the TNF inhibitor does not assume its proper, active structure, any disulfide bonds which have formed and/or any noncovalent interactions which have occurred will first be disrupted by denaturing and reducing agents, for example, guanidinium chloride and beta-mercaptoethanol, before the TNF inhibitor is allowed to assume its active structure following dilution and oxidation of these agents under controlled conditions.

For purifications prior to and after refolding, some combinations of the following steps is preferably used; anion exchange chromatography (monoQ or DEAE-Sepharose), gel filtration chromatography (superose), chromatofocusing (MonoP), and hydrophobic interaction chromatography (octyl or phenyl sepharose). Of particular value will be affinity chromatography using TNF (described in Example 1).

(b) TNF inhibitor Produced from Mammalian Cells.

TNF inhibitor produced from mammalian cells will be purified from conditioned medium by steps that will include ion exchange chromatography and affinity chromatography using TNF as described in Example 1. It will be apparent to those skilled in the art that various modifications and variations can be made in the processes and products of the present invention. Thus, it is intended that the present invention cover the modifications and variations of this invention provided they come within the scope of the appended claims and their equivalents.

As indicated previously, the TNF inhibitors of the present invention are contemplated for use as therapeutic agents and thus are to be formulated in pharmaceutically acceptable carriers. In one embodiment of the present invention, the TNF inhibitors may be chemically modified to improve the pharmokinetic properties of the molecules. An example would be the attachment of the TNF inhibitors to a high molecular weight polymeric material such as polyethylene glycol. In addition, interleukin-1 inhibitors may be administered in conjunction with the TNF inhibitors. This combination therapeutic will be especially useful in treatment of inflammatory and degenerative diseases.

The following examples illustrate various presently preferred embodiments of the invention claimed herein. All papers and references cited in the Examples that follow are specifically incorporated herein by reference.

Example 1. Protein Preparation

A. Materials

The gene for TNF alpha (TNFa) was purchased from British Biotechnology, Limited, Oxford, England. DEAE-Sepharose CL-6B resin and Mono-Q HR5/5, HR10/10 FPLC columns were purchased from Pharmacia, Inc., Piscataway, New Jersey. Affigel-15 resin, and BioRad protein assay kit were purchased from BioRad, Richmond, California., Tween 20, ammonium bicarbonate, sodium phosphate, PMSF, sodium bicarbonate, dithiothreitol crystal violet and actinomycin D were purchased from Sigma Chemical Company, St. Louis, Missouri. Endoproteinase Lys-C, Endoproteinase Asp-N and TRIS were purchased from Boehringer Mannheim Biochemicals, Indianapolis, Indiana. Hexafluoroacetone was purchased from ICN Biomedicals, Costa Mesa, California. Cyanogen bromide, trifluoroacetic acid, and guanidine hydrochloride were purchased from Pierce Chemicals, Rockford, Illinois.

Acetonitrile and HPLC water were purchased from J.T. Baker Chemical Company, Phillipsburg, New Jersey. Urea was purchased from Bethesda Research Laboratories, Gaithersburg, Maryland. [^{3}H]-Iodoacetic acid was purchased from New England Nuclear, Boston, Massachusetts. [^{125}I]-TNFa was purchased from Amersham, Arlington Heights, Illinois. Recombinant human TNFa was purchased from Amgen, Thousand Oaks, California. C8-reverse phase columns (25 cm x 4.6 mm) were obtained from Synchrom, Inc., Lafayette, Indiana. A C8-microbore reverse phase column (7 micron, 22 cm x 2.1 mm) was obtained from Applied Biosystems, Foster City, California. Corning 96-well microtiter plates were purchased from VWR Scientific, Batavia, Illinois. McCoys 5A media and fetal bovine serum were purchased from Gibco, Grand Island, New York. RPM-1 1640 media and L-glutamine were purchased from Mediatech, Herndon, Virginia. Trypsin was purchased from K. C. Biologicals, St. Lenexa, Kansas. ME180, U937 and L929 cell lines were obtained from American Type Culture Collection, Rockville, Maryland.

B. Assays for the TNF inhibitor

Two types of assays were used to identify the TNF inhibitor. One of them is a cytotoxicity assay. The other is a gel shift assay.

1. Cytotoxicity Assay

The cytotoxicity assay was performed with actinomycin D-treated ME180 cells and L929 cells as described by Ostrove and Gifford (Proc. Soc. Exp. Biol. Med. 160, 354-358 (1979)) and Aggarwal and Essalu (J. Biol. Chem. 262, 10000-10007 (1987)). L929 cells (CCLI: American Type Culture Collection) cells were maintained in McCoy's 5A medium containing 10% fetal bovine serum. Confluent cultures were treated briefly with 0.25% trypsin in physiological solution containing 5mM EDTA and resuspended in a fresh medium. Approximately 2 x 10^{4}

trypsinized cells per well were plated in 96-Well plates (Corning) and incubated for 24 hours at 37°C. Then actinomycin D was added to a final concentration of 0.25 ug per ml. After two hours, samples containing TNF and TNF inhibitor were added to the wells and incubation was continued overnight at the same temperature. After microscopic evaluation, the medium was decanted, and the wells were rinsed with PBS. The wells were then filled with a solution of 0.1% crystal violet, 10% formaldehyde and 10mM potassium phosphate, pH 6.0 for 5 min, washed thoroughly with water and dried. The dye was extracted with 0.1M sodium citrate in 50% ethanol, pH 4.2. The absorbance of the dye in viable cells was determined at 570 nm using a Kinetic microplate reader (Molecular Devices Corp. CA). An example of this assay is shown in Figure 1. In the presence of TNF inhibitor, the cytotoxic effect of TNF was reduced.

2. Gel Shift Assay

The gel shift assay involves the use of a native polyacrylamide gel electrophoresis system. This native 4% gel electrophoresis was performed according to Hedrick and Smith (Arch. Biochem. and Biophysics 126, 155-164 (1968)). The iodinated TNF (Amersham) was mixed with the TNF inhibitor from Example 1.C. after C8 chromatography and incubated for 30 min. to 2 hours. This mixture, along with the iodinated TNF alone, were loaded onto the 4% native gel and electrophoresed. After the gel was fixed with 10% acetic acid and washed, a film was placed for radioautography. As shown in Figure 2, the complex of TNF and TNF inhibitor migrates differently from the TNF by itself. This gel shift assay was used to determine which fractions contain TNF inhibitor in the eluates of DEAE CL6B column chromatography.

C. Purification of the 30kDa TNF Inhibitor

Twenty liters of urine from a patient diagnosed with renal dysfunction was concentrated to 200 ml with an

Amicon YM5 membrane. The concentrate was then dialyzed at 4°C against 0.025 M Tris-Cl, pH 7.5, and subsequently centrifuged in a JA14 rotor at 10,000 rpm for 30 minutes. The supernatant was then loaded onto a 40 x 4.5 cm DEAE Sepharose CL-6B column equilibrated with 0.025 M Tris-Cl, pH 7.5 and extensively rinsed with equilibration buffer until the OD_{280} of the effluent returned to baseline. Chromatography was accomplished using a linear gradient from 0-0.05 M sodium chloride in 0.025M Tris-Cl pH 7.5 and monitored by OD_{280}. Column fractions were collected, and assayed for TNF inhibitor activity using the native gel assay. The TNF inhibitor eluted elutes in a rather sharp peak at 80mM NaCl.

Figure 6A shows the OD_{280} profile of the DEAE Sepharose CL-6B chromatography of 20 1 urine. Figure 6B shows the autoradiograph of the corresponding native gel assay indicating a peak of the TNF inhibitor at fractions 57-63, which is about 80mM NaCl.

The TNF inhibitor was further purified using a TNF affinity column. Recombinant TNF was expressed in BL21/DE3 at about 10-20% total cell protein. The cell pellet was French-pressed at 20,000 psi and the soluble material dialyzed at 4°C against 0.025 M Tris-Cl pH 8.0. The dialyzed lysate was 0.2 micron-filtered and loaded onto a Mono-Q FPLC column equilibrated with 0.025 M Tris-Cl ph 8.0. A linear gradient from 0 to 0.5 M NaCl in 0.025 M Tris-Cl pH 8.0 was run and monitored by OD_{280}. One ml fractions were collected and analyzed for purity by SDS-PAGE. The subsequent TNFa pool was about 90% pure based on Coomassie-stained SDS-PAGE and was fully active based on a Bradford protein assay, using lysozyme as a standard, and an ME180 bioassay, using Amgen's TNFa as a standard (Bradford, M. Annal. Biochem. 72, 248-254 (1976)).

TNFa was concentrated in an Amicon Centriprep-10 to about 25 mg/ml, dialyzed against 100 mM $NaHCO_3$, pH 8.5, and coupled to Affigel-15 resin at 25 mg TNF/ml resin. A

coupling efficiency of greater than 80% yielded a high capacity resin which was used to further purify the TNF inhibitor.

PMSF, at a final concentration of 1-4 mM, was added to the DEAE CL-6B pool and applied to a 4 x 1 cm TNF affinity column equilibrated at 4°C with 0.025 M Tris-Cl pH 7.5 at a flow rate of 0.1 ml/min. The column was then rinsed with 0.025 M Tris-Cl pH 7.5 until the OD_{280} of the effluent returned to baseline. The column was subsequently eluted with 0.05 M NaPhos, pH 2.5 and monitored by OD_{280}. Figure 7 shows the OD_{280} profile of the 0.05 M NaPhos pH 2.5 elution from the TNF affinity column.

The TNF inhibitor was purified to homogeneity by reverse phase HPLC on a Syncropak RP-8 (C8) column. The OD_{280} peak from the TNF affinity column was pooled and immediately loaded onto a RP-8 column, equilibrated with 0.1% TFA/H_2O, a linear 1%/min gradient of 0.1% TFA/acetonitrile was run, from 0-50%, and monitored by OD_{215} and OD_{280}. Fractions were collected and assayed from bioactivity using L929 cells and the native gel assay described in Example 1.B. Both of these assays indicate bioactivity at fractions 28-32 which corresponds to a peak of OD_{215} and OD_{280} eluting at 18% acetonitrile.

Figure 8A and 8C show the chromatographic profile of the TNF affinity pool on a Syncropak RP-8 column with the corresponding bioactivity from the L929 cytotoxicity assay. Figure 8B shows a silver stained 15% reducing SDS-PAGE of the RP-8 pool indicating a single band at 30kDa.

D. Characterization of the Protein Component of 30kDa TNF Inhibitor

30kDa TNF inhibitor is a glycoprotein as was detected using Concanavalin A-Peroxidase after the protein was transferred onto the nitrocellulose filter. This method is a modification of Wood and Sarinana (Analytical

Biochem. 69, 320-322 (1975)) who identified glycoproteins on an acrylamide gel directly. The peroxidase staining of glycoprotein was performed by using peroxidase conjugated Con A or non-conjugated Con A. When non-conjugated Con A was used, the nitrocellulose filter was incubated for one hour in a solution containing Con A (0.5 mg/ml, Miles Laboratory) in phosphate buffer, pH 7.2 (PBS); then washed 3 x 5 min. in PBS. The washed filter was incubated in horseradish peroxidase (0.1 mg/ml, Sigma Chemical) for one hour. After 3 x 15 min. wash in PBS the filter was immersed in a solution containing 3 mg/ml 4-chloro-1-naphthol (Sigma Chemical) and 12.5 ul/ml of hydrogen peroxide until the color was developed. Glycoprotein was seen as a purple color. A photograph was made as soon as the filter was developed as shown in Figure 3.

Chemical deglycosylation of TNF inhibitor was carried out by the method of Edge, Faltynek, Hof, Reichert and Weber (Analytical Biochem. 118, 131-137 (1981)). A mixture of 0.25 ml anisole (Eastman Kodak) and 0.5 ml of trifluoromethanesulfonic acid (Eastman Kodak) was cooled to 4°C, then 1-200 ng of dry TNF inhibitor were dissolved in 3ul of this mixture. The tube was flashed with nitrogen, then incubated for 30 min. at room temperature. This deglycosylated protein was analyzed on SDS-PAGE (Figure 4). The molecular weight of chemical treated TNF inhibitor is about 18,000 dalton. A band at 14,000 was seen also, but this may be a proteolytic fragment of deglycosylated TNF inhibitor.

The enzymatic deglycosylation using N-glycanase was performed following the manufacturer's protocol (Genzyme Corp.) except TNF inhibitor was incubated with N-glycanase for 5 to 6 hours instead of overnight. The molecular weight of the deglycosylated form of denatured TNF inhibitor is shown to be about 20,000 dalton (Figure 5). When the inhibitor is not denatured prior to deglycosylation, the molecular weight of the

deglycosylated protein is about 26,000 dalton.

E. Deglycosylated 30kDa TNF-inhibitor binds to TNF.

Radiolabeled TNF inhibitor (30kDa) was treated with TFMSA (trifluoromethanesulfonic acid) in order to remove carbohydrates, and the TFMSA was separated from the protein by HPLC. The protein fraction was mixed with TNF-affigel for one hour at 4° C, and all unbound material was removed by centrifugation. The TNF-affigel was washed extensively with 50mM NaP04, pH 2.5. Radioactivity in each fraction was counted and also analyzed on a SDS-PAGE. Non-specific binding of TNF inhibitor was measured using anhydrochymotrypsin affigel. The results are shown in Table 2. These results indicate that deglycosylated TNF inhibitor (30kDa) binds to TNF.

TABLE 2

Sample	Type of Affinity	Count (CPM) Flow Through	Eluate
Native TNF-INH	TNF	49401 (55.0%)	40014 (45.0%)
Native TNF-INH	Anhy CT	80000 (98.0%)	1789 (2.0%)
TFMSA-Treated TNF-INH	TNF	13369 (73%.0)	4908 (27.0%)
TFMSA-Treated TNF-INH	Anhy CT	15682 (94.0%)	926 (6.0%)

In another experiment, radiolabeled TNF inhibitor (30kDa) was reduced, then deglycosylated with N-glycanase. After deglycosylation, the material was incubated with 13 mM oxidized glutathione (GSSG) for 10 minutes at room temperature, and diluted 5 fold with 50mM Tris. Cysteine was then added to a final concentration of 5mM. The material was incubated at 4° C for 16 hours then mixed with a TNF-affigel for one hour at 4° C. Unbound material was removed, and the gel was washed extensively with 50mM Tris-HC1, pH 7.5. The bound material was eluted with 50mM $NaPO_4$, pH 2.5. Radioactivity in each fraction was analyzed, and a SDS-

PAGE was performed for each fraction. As seen in Table 3 and Figure 18, the deglycosylated and reoxidized TNF inhibitor also binds to TNF.

TABLE 3

Sample	Type of Affinity	Count (CPM) Flow Through	Eluate
Native TNF-INH	TNF	18281 (60.0%)	12603 (40.0%)
Native TNF-INH (reduced/reoxidized)	TNF	28589 (94.0%)	1964 (6.0%)
TFMSA-Treated (reduced/reoxidized)	Anhy CT	31371 (98.70)	421 (1.3%)
Deglycosylated TNF-INH (reduced/reoxidized)	TNF	25066 (85.0%)	4305 (15.0%)
Deglycosylated TNF-INH (reduced/reoxidized)	Anhy CT	29619 (98.4%)	495 (1.6%)

Example 2. Sequencing of 30kDa TNF Inhibitor

N-terminal sequences were determined using Applied Biosystems Protein Sequencers, models 470 and 477. Prior to sequencing, peptides generated from a variety of proteolytic enzymes were purified on an Applied Biosystems C8-microbore HPLC column (22 cm x 2.1 mM).

A. Amino Terminal Sequencing

Approximately 250 pmoles of reverse phase (RP-8) purified TNF inhibitor were applied directly to a polybrene filter and subjected to automated Edman degradation. The resulting sequence information yielded the first 30 amino acids of the molecule.

B. Endoproteinase Lys-C Digestion of Native Protein

Approximately 250 pmoles (5 ug) of reverse-phase purified TNF inhibitor was digested with 1 ug of endoproteinase Lys-C. The 12 hour digestion at 25°C was carried out in the presence of 1M urea, 0.01% Tween 20, and 150 mM NH_2HCO_3, pH 8.0. Prior to peptide purification the digest was reduced by incubation for 1 hour following addition of 50-fold molar excess of dithiothreitol, or

reduced and alkylated by a further one hour incubation at 37°C using a two-fold molar excess of [^{3}H]-iodoacetic acid over dithiothreitol. Figure 9A shows the reverse phase HPLC pattern of this digestion. Figure 9B shows the reverse phase HPLC pattern of this digest followed by alkylation.

C. Endoproteinase Asp-N Digestion of Native Protein

Approximately 250 pmol (5ug) of reverse phase purified TNF inhibitor was digested with 0.5-2.5 ug endoproteinase Asp-N. The 12-18 hour digest at 37°C was carried out in the presence of 1M guanidine-HCl, 0.01% Tween 20 and 150 mM NaPhos, pH 8.0.

Prior to peptide purification the digest was reduced and alkylated as in Example 2.B. Figure 10 shows the reverse phase HPLC pattern of two such digests.

D. Reduction Carboxymethylation of Protein

The reverse-phase HPLC purified TNF inhibitor was reduced and carboxymethylated with [^{3}H] Iodoacetic acid as described by Glazer, et al., in Chemical Modifications of Proteins, pp. 103-104 (1975), except two successive rounds of reduction followed by alkylation were used. The protein was re-purified by reversé-phase HPLC prior to proteolytic digestion.

E. Endoproteinase V8 Digestion of Reduction Carboxymethylation of Protein

An analytical digest was performed by dissolving 55 pmoles (about 1 ug) of reduced carboxymethylated TNF inhibitor in 150 mM $NaHCO_3$ pH 8.0, and digesting it with 0,2 ug V8 protease for 18 hours at 25°C. Reverse-phase HPLC (Figure 11A) revealed three sequenceable peptides and indicated a larger scale digest was in order. Approximately 220 pmoles (4.5 ug) of reduced carboxymethylated TNF inhibitor was digested with 1 ug V8 protease for 5 hours at 25°C., when an additional 0.5 ug V8 protease was added and the digestions continued for 16 hours. Figure 11B shows the reverse-phase HPLC of the

large scale V8 digest.

F. Complete Primary Structure of 30kDa TNF Inhibitor Based on Peptides Sequences and cDNA Sequence.

Various peptide fragments were aligned according to the cDNA sequence obtained in Example 4. This is shown in Figure 19. Residues which are not identified by protein sequencing are residue numbers 14, 42, 43, 44, 96, 97, 105, 107, 108, and 110 through 119. The sequence of Gln-Ile-Glu-Asn is apparently the carboxyl terminus of the 30 kDa TNF inhibitor.

Example 3. 30kDa TNF inhibitor is produced by U937 cells stimulated with PMA and PHA.

The monocyte-like cell line U937 was grown at 37°C in RPMI medium containing 10% fetal calf serum to a cell density of 1 x 10^6 cells/ml. The cells were then removed by centrifugation and resuspended on 5 different 100 cm^2 petri plates at 2 x 10^6 cells/ml in RPMI without serum containing 10 ng/ml of PMA (phorbol 12-myristate 13-acetate) and 5 ug/ml PHA-P (phytohemagglutinin-P). The conditioned medium from one plate was harvested after only 10 minutes of incubation and used as a zero time control. The medium from the remaining plates was successively removed at 24 hours, 48 hours, 72 hours and 96 hours after plating. The protein contained in these samples was concentrated into approximately 400 ul each by Centriprep-10 (Amicon Corp.) treatment. Each 400 ul sample was then mixed with an equal volume of an Affigel-15 (Biorad Corp.) preparation containing approximately 10 mg/ml of purified human recombinant TNFa that had been prepared in our laboratory. This TNFa, prior to being bound to the Affigel-15 resin, had been shown to be bioactive by its toxicity to murein L929 cells.

The conditioned medium was incubated at room temperature batchwise with the TNFa affinity resin for 2

hours. The unbound fraction was removed after centrifugation of the resin and the resin was subsequently washed with 1 ml (500ul, 2x) of PBS (phosphate buffered saline, pH 7.5) containing 0.1% gelatin. Bound material was eluted with a 25 mM solution of monobasic sodium phosphate, pH 2.5 (400 ul, 2x). 40 ul of each of the unbound, washed, and eluted fractions were dried, resuspended in 10 ul of 25 mM Tris pH 7.5, mixed with 2 ul (100 pci) of ^{125}I-TNFa (400-800 ci/mmole, Amersham) and incubated for 30 minutes at room temperature. These mixtures were then mixed with 5 ul of 40% sucrose and 1 ml of 0.1% bromophenol blue and applied to a 4% native acrylamide gel as described in Example 1.B. The conditioned medium from all samples except the zero control contained TNFa binding activity by this assay as shown in Figure 15.

The remaining 300 ul from each sample (1st low pH elution) were applied to a C8 HPLC column and eluted with a linear gradient of acetonitrile over 60 minutes (1%/minute, 1 ml/minute flow rate, 1 ml fractions were collected). Each fraction as dried and resuspended in 50 ul of PBS + 0.1% gelatin. 10 ul of each of these samples was mixed with ^{125}I-TNFa as above and analyzed by native polyacrylamide gel. TNFa binding activities are detected in fractions corresponding to 33% and 36% acetonitrile as shown in Figure 16.

Example 4. Analysis of messenger RNA from PMA/PHA treated U937 cells

U937 cells were grown as described in Example 3 to a density of 1 x 10^6 cells/ml and then resuspended in serum-free medium at 2 x 10^6 cells/ml without or with PMA (10 ng/ml) and PHA (5 ug/ml). Samples were taken at 1 hour +/- PMA/PHA and 17 hours + PMA/PHA only. Total RNA was prepared from the cells by the guanidinium thiocyanate-phenol-chloroform method of Chomczynski and Sacci (Analytical Biochemistry 162:156-159, (1987)).

Poly A^+ RNA was prepared from total RNA by annealing to oligo dT cellulose (Bethesda Research Labs). Eight micrograms of each poly A^+ RNA was then applied to a 6.6 formaldehyde, 1.2% agarose gel. The RNA within the gel was then blotted to a zeta probe membrane (BioRad). The membrane was treated as described in Example 5 for screening of a human genomic DNA library with oligonucleotide probes. 1×10^6 cpm/ml of a labelled single stranded DNA probe (polynucleotide kinase) was added. The sequence of this probe is:

5' TTGTGGCACTTGGTACAGCAAAT 3'

and it corresponds to bases 410-433 of the sequence set forth in Figure 13. Following overnight hybridization at 65°C, the membrane was washed once at room temperature in 6 X SSC 0.1% SDS and once at 65°C in the same solution and then exposed to x-ray film for 72 hours. The autoradiogram shown in Figure 17 shows that PMA/PHA treatment of U937 cells in serum-free medium for 1 hour clearly stimulates the expression of the 30kDa TNFa inhibitor messenger RNA and that by 17 hours of treatment this message is virtually absent from the cells. The molecular size of the 30kDa TNFa inhibitor messenger RNA based on this experiment is approximately 2.4 kilobases.

Example 5. Preparation of a human genomic DNA library for 30kDa TNF inhibitor

Human genomic DNA was partially digested with Sau3AI and size selected. DNA with an average size of 15 KB was ligated into the BamHI site of bacteriophage lambda Charon 30. (Rimm, D.L., Horness, D., Kucera, J., and Blattner, F.R. Gene 12:301-309 (1980)). Phage were propagated and amplified on E. coli CES 200.

A. Probes

The four degenerate oligonucleotide hybridization probes listed in Table 4, were synthesized on an Applied Biosystems DNA synthesizer. Each probe mixture consisted of all possible DNA sequences coding for the given

peptide sequence.

TABLE 4

Peptide Name	Peptide Sequence	Probe Name	Probe Sequence
LysC 18	KEMGQVE	TNFBP-P20	5'TCNACTCTGNCCCATTCTCTCTT 3'
LysC 11	QGKYIHP	TNFBP-P2'	5'CAAGGGNAAAGTATCACATCC 3'
LysC 11	YNDCPG	TNFBP-P3'	5'TATCAATCGATCTGTCCCNGG 3'
LysC 11	YIHPQNN	TNFBP-P4	5'TTAGTTTCTGNGGAGTCAGT 3'

N = G, A, T, or C.

Oligonucleotides were labeled with [gamma $-_{32}$P] ATP (Amersham Inc., Arlington Heights, IL) and T4 polynucleotide kinase (Boehringer Mannheim, Indianapolis, IN) to a specific activity of 6-9 x 10^6 c.p.m./picomole according to manufacturer's instructions.

B. Methodology:

8.4 x 10^5 lambda phage containing human genomic DNA were plated and transferred to duplicate nitrocellulose filters. These filters were hybridized with 1 pMol/ml of probe TNFBP-P2' for 16 hours in a solution containing 1.0 M NaCl, 0.1 M sodium citrate, 2x Denhardts solution (Denhardt, D.T. Biochem. Biophys, Res. Commun. 23:641-646 (1966)), 0.1% SDS, 0.05% sodium pyrophosphate and 150 ug/ml yeast tRNA at a temperature of 52°C. This temperature is 2°C below the calculated Tm for the most AT-rich member of the oligonucleotide pool. (Suggs, S.V. in Developmental Biology Using Purified Genes, (Brown, D.D., and Fox, C.F., eds.) Academic Press, New York, pp. 683-693 (1981)). After hybridization, the filters were washed for 45 minutes at ambient temperature with three changes of 1 M NaCl, 0.1 M sodium citrate and 0.5% SDS. A stringent wash of eight minutes was done at the calculated Tm (i.e., 2°C above hybridization temp) for the most AT-rich member to the pool. Filters were then

dried and autoradiographed for 40 hours with one intensifying screen at -70°C.

Eleven positive hybridizing plaques were detected and these were isolated and amplified. The ability of these clones to hybridize to TNFBP-P20, TNFBP-P3' and TNFBP-P4 was tested using similar methodology. One clone (TNFBP-8) hybridized to all four oligonucleotides. This clone was plaque purified and amplified. DNA was prepared from this clone using Lambda-Sorb (Promega Corporation, Madison, WI) and a method described by the manufacturer.

One microgram of this DNA was then digested with Sau3AI and the fragments subcloned into BamHI digested M13 sequencing vector mp 18 (Yanish-Perron, C., Viera, J., and Messing, J. Gene 33:103-119 (1985)). M13 clones were then transferred to duplicate nitrocellulose filters and hybridized to the oligonucleotide probes in Table 4 using conditions previously described. Positive subclones were purified and sequenced (Sanger, F., and Coulson, A.R.J. Mol. Biol. 94:441-448 (1975)) using a modified T4 DNA polymerase (Sequenase, US Biochemical Corp., Cleveland OH) as described by the manufacturer, and using as primers either the degenerate probes used to identify the clone or sequence obtained using those probes. Among the sequences obtained are those of Subclones TNFBP-M13-Sau3A-P2'-2 and TNFBP-M13-Sau3A-P4 Primers P3, P3', P2', P2 and P4. The sequence data is set forth in Figure 13. The sequence contains DNA coding for at least 48 amino acids of 30kDa TNF inhibitor peptides other than those specified by the probes and therefore confirms that the clone TNFBP8 codes for TNF inhibitor. The sequence also shows that the gene for TNF inhibitor includes at least one intron (GTAGGGGCAA . . . . . .CCCCATTCACAG). Finally, this sequence shows that 30kDa TNF inhibitor is synthesized as a precursor protein and that a proteolytic cleavage at the Arg-Asp sequence is required to generate the mature, active protein.

Example 6. Preparation and screening of a cDNA library of mRNA from U937 cells stimulated with PMA/PHA.

The experiment described in Example 4 shows that U937 cells treated with PMA/PHA for 1 hour should contain a pool of messenger RNA enriched for the TNF inhibitor (30kDa). Accordingly, a cDNA library was prepared from polyA$^+$ RNA obtained from U937 cells treated with PMA/PHA as described in Example 4. Double stranded, blunt ended cDNA was obtained from approximately 5 ug of poly A$^+$ RNA essentially as described by Gubler, U., and Hoffman, B.J., (1983 Gene, 25:263) using lot tested reagents (Amhersham, Arlington Heights, IL) according to procedures recommended by the manufacturer. Approximately 1 ug of double stranded cDNA obtained was treated with the enzyme EcoRI methylase and EcoRI linkers having the sequence: d(pCCGGAATTCCGG) (New England Biolabs, Beverly, MA), were attached via T4 DNA ligase followed by digestion with endonuclease EcoRI. This DNA was then ligated into a lambda-bacteriophage cloning vector gt10 (Young, R.A., and Davis, R.W. (1983) Proc Natl Acad Sci USA, 80:1194-1198) that had been digested with EcoRI and the product packaged into infective lambda-bacteriophage particles using lambda-DNA packaging extracts (Gigapack II Gold) obtained from Stratagene (La Jolla, CA) according to their protocol. This lambda-lysate (cDNA library) was then used to infect E. coli strain C600 hflA and it was shown that the library contained approximately 2.5 x 10^6 recombinant members.

Approximately 4 x 10^5 members of this library were plated on E. coli strain C600 hflA (5 x 10^4 p.f.u./plate). Duplicate lifts to nitrocellulose were made and the filters were treated as described in Example 5 for screening of the human genomic library. The DNA on the filters was then hybridized to the same ^{32}P labelled probe as described in Example 4 except that the temperature of incubation was 42°C. From 4 x 10^5 recombinant phage plated, 3 duplicate plaques hybridized

to this probe. These were further reisolated and probed as above and with an additional synthetic probe having the sequence:

5' CCCCGGGCCTGGACAGTCATTGTA 3'

This probe corresponds to bases 671-694 of the human genomic TNF inhibitor clone shown in Figure 13. Both probes hybridized to all three plaques identified with the first.

After plaque purification DNA was prepared from these three clones and subcloned into the EcoRI site of M13 vectors MP18 and MP19 as described in Example 5. Each of these cDNAs consists of two EcoRI fragments one of approximately 800 bp common to all three clones and another 1300 bp, 1100 bp or 1000 bp depending on the clone. The likely origin of the unique EcoRI fragments in each clone is incomplete elongation by the enzyme reverse transcriptase during 1st strand synthesis of the cDNA. Therefore, those EcoRI fragments likely represent the 5' end of the TNF inhibitor mRNA and the 800 bp fragment the 3' end. This is confirmed by the DNA sequence obtained for these fragments as described below.

From the EcoRI subclones of the cDNA described above the entire sequence of the 2100 bp cDNA was obtained. The dideoxy nucleotide chain termination method of sequencing was used (Sanger, F. and Coulson, A.R. (1975) J. Mol. Biol. 94:441-448). The modified T7 DNA polymerase, Sequenase (U.S. Biochemical, Cleveland, OH) was used as the elongation enzyme as described by the supplier. Sequencing primers were synthetic oligonucleotides prepared from the human genomic sequence of the TNF inhibitor as shown in Figure 13 or sequences obtained using those primers. Figure 20 shows the translated sequence derived from one of the cDNA clones. This sequence corresponds to that obtained by protein sequence data as described in Figure 19. The entire sequence of the human 30kDa TNF inhibitor cDNA from clone lambda-gt10-7ctnfbp is shown in Figure 21.

Example 7. Expression of the 30kDa TNF inhibitor cDNA in Escherichia coli

The portion of the TNF inhibitor (30kDa) cDNA gene coding for the soluble TNFa binding activity has been prepared for expression in E. coli as described below.

Because the protein coding sequence defining the C-terminal portion of the urine derived TNF inhibitor (sequence QIEN, base 771 Figure 20) is not followed by a termination codon in the cDNA sequence, one was added by oligonucleotide directed in vitro mutagenesis (Biorad, Richmond, CA). An M13MP19 clone of the 1300 bp EcoRI fragment from the clone lambda-gt107ctnfbp, was hybridized with the synthetic oligonucleotide:

5' CTACCCCAGATTGAGAATTAAGCTTAAGGGCACTGAGGAC 3'

After 2nd strand synthesis and transfection into an appropriate host, mutant clones were identified by hybridization to the above described mutagenic oligonucleotide. The molecular identity of the clones so identified was confirmed by DNA sequencing as described (Example 5). Next, a 468 bp fragment defined by StyI (position 303) and HindIII defining the C-terminus of the protein was removed from the Rf form as a mutagenized clone and inserted into E. coli expression plasmid containing the tacI promoter (DeBoer, H.A., et al., (1983) Proc. Natl. Acad. Sci. USA 80:21-25). This construction was accomplished by use of the synthetic, double strand adapter sequence:

5' GATCCGATCTTGGAGGATGATTAAATGGACAGCGTTTGCCCC 3'

GCTAGAACCTCCTACTAATTTACCTGTCGCAAACGGGGGTTC

This adapter translationally couples the TNF inhibitor gene (truncated form as described above) to the first 12 codons of the bacteriophage T7 gene 10. The DNA sequence of this construct from the point of translation initiation at gene 10 through the adapter sequence is shown in Figure 22. A methionine codon (ATG) is necessarily added to the TNF inhibitor gene sequence for

expression in E. coli. This plasmid is called pTNFiX-1.

The predicted molecular weight of this protein is approximately 17,600kDa a molecular weight that is very close to the deglycosylated native TNF inhibitor (30kDa).

Example 8: Purification of active TNF inhibitor (30kDa) from Escherichia coli

Cells from one liter of E. coli culture (pTNFIX-1JM1071on-) grown under induced condition for 2 hours were resuspended in 10 ml of 50mM Tris-HCl, pH 7.5 containing 2mM EDTA (TE buffer) and French pressed at 20,000 psi. at 4°C. The material was centrifuged at 20,000g for 10 min. The resulting pellet was washed once with TE-buffer. The washed pellet was resuspended in 2 ml of 6M Guanidine-HCl and incubated at room temperature for 10 min. After the incubation, 80 ul of 500 mM DTT was added and the mixture was incubated at room temperature for another 30 min. The material which remained insoluble after this treatment was removed by centrifugation at 20,000g for 15 min. 120 ul of 500 mM oxidized glutathione was added to the supernatant, and the mixture was incubated at room temperature for 10 min. This material was then diluted in 20 ml of 0.6% Tris base solution, and 220 ul of 500 mM cysteine was added. The incubation was continued for another 16 hours at 4°C. After 16 hours of incubation, some insoluble residue was observed. This insoluble material was removed by centrifugation at 20,000g for 20 min. The resulting supernatant was dialyzed against 50mM Tris-HCl pH 7.5 for 16 hours at 4°C., then centrifuged at 20,000g for 10 min. PMSF at a final concentration of 4mM was added to this supernatant and this material was loaded onto a TNF-affinity column (.7 x 2cm) at a flow rate of 0.1 ml per min. This column was extensively washed with 50mM Tris-HCl pH 7.5, and bound proteins were eluted with 50mM $NaPO_4$-HCl pH 2.5 The pH 2.5 eluate was loaded onto an RP8 column which was previously equilibrated with 0.1%

TFA/H20. TNF inhibitor was eluted with a linear gradient of 0.1% TFA/acetonitrile at 1%/min. (Figure 25). Fractions were analyzed on SDS-PAGE (Figure 26), and cytotoxicity assay was performed (Figure 25) to localize the TNF inhibitor. The E. coli-produced TNF inhibitor (30kDa) migrates to about 20 kDa, since it is not glycosylated. Fractions number 30 through 35 contain TNF inhibitor. The amino terminal sequence of this material shows that the E. coli produced TNF inhibitor has the following sequence:

Met-Asp-Ser-Val-()-Pro-Gln-Gly-Lys-Tyr-Ile-His-Pro-Gln-Asn-Asn-Ser-

By using this procedure, about 40 ug of TNF inhibitor (30kDa) was obtained from one liter of the culture. The yield was about 2 to 3%. The yield can be increased to over 50% by purifying the TNF inhibitor before refolding.

Example 9. Expression of genes encoding 30kDa TNF inhibitor in animal cells

Animal-cell expression of TNF inhibitor requires the following steps:

a. Construction of an expression vector.

b. Choice of host cell lines.

c. Introduction of the expression vector in host cells.

d. Manipulation of recombinant host cells to increase expression levels of TNF-BP.

1. TNF inhibitor expression vectors designed for use in animal cells can be of several types including strong constitutive expression constructs, inducible gene constructs, as well as those designed for expression in particular cell types. In all cases, promoters and other gene regulatory regions such as enhancers (inducible or not) and polyadenylation signals are placed in the appropriate location in relation to the cDNA sequences in plasmid-based vectors. Two examples of such constructs follow.

A construct using a strong constitutive promoter region can be made using the cytomegalovirus (CMV) immediate early gene control signals. This plasmid can be constructed using standard molecular biological techniques (Maniatis, et al., Molecular Cloning, a Laboratory Manual. Cold Spring Harbor Laboratory, 1982) and resulting in the plasmid shown in Figure 23.(pCMVXV beta TNFBPstopA) The SV40 origin of replication is included in this plasmid to facilitate its use in COS cells for transient expression assays. This particular construct contains the CMV immediate early promoter and enhancer as described by Boshart, et al., (Cell 41:521-530, 1985) followed by the rabbit B-globin second intron (see van Ooyen et al., Science 206:337-344, 1979) which is flanked by BamHI and EcoRI restriction sites. This intron is included because expression levels have been shown to be increased when introns are included in the transcribed regions of some expression vectors (Buckman and Berg, Mol. Cell. Biol. 8:4395-4405, 1988). The polyadenylation signal is provided by simian virus 40 (SV40) sequences (map coordinates 2589-2452; see Reddy, et al., Science 200:494-502, 1978). The 30kDa TNF inhibitor cDNA sequences have been modified as follows: the extensive region located 3' of the C-terminus of the purified TNF inhibitor from human urine has been deleted and a stop codon has been engineered into the position just following the C-terminal asparagine. The unmodified 30kDa TNF inhibitor cDNA sequences in an analogous vector have been inserted into COS cells and been shown to increase the TNF binding activity of such cells.

The second construct (see Figure 24) (pSVXVTNFBP stop A) uses the strong constitutive promoter region from the SV40 early gene in an arrangement such as that found in the plasmid pSV2CAT (Gorman, et al., Mol. Cell. Biol. 2:1044-1051, 1982). This plasmid should be manipulated in such a way as to substitute the TNF inhibitor cDNA for the chloramphenicol acetyltransferase coding sequences

using standard molecular biological techniques. Once again, the TNF inhibitor cDNA has been modified as described above for the CMV promoter construct. The SV40 early promoter region includes sequences from the HindIII site to the BamHI site (map coordinates 5090-188; see Reddy et al., Science 200:494-502, 1978) and the SV40 polyadenylation signal is as described above for the CMV construct.

2. Two animal cell lines have been used to express TNF inhibitor using the vectors described above to produce active protein. Cell lines that have been characterized for their ability to promote expression of this foreign gene include the monkey kidney cell, COS-7, and Chinese hamster ovary (CHO) dihydrofolate reductase deficient (dhrf-) cells.

3. To establish a continuous CHO-derived cell line that secretes 30kDa TNF inhibitor into cell culture medium, a TNF inhibitor expression plasmid has been introduced into these dhfr- cells along with a plasmid that directs the synthesis of dihydrofolate reductase using the calcium phosphate-DNA precipitation technique described by Graham and van der Eb (Virology 52:456-467, 1973). The cells that have taken up DNA and express DHFR were selected as described by Ringold, et al., (J. Mol. Appl. Genet. 1:165-175, 1981).

4. Cells that express the TNF inhibitor gene constructs can be manipulated to increase the levels of production of TNF inhibitor. Cells containing TNF inhibitor expression vectors along with a dhfr expression vector should be taken through the gene amplification protocol described by Ringold, et al., (J. Mol. Appl. Genet. 1:165-175, 1981) using methotrexate, a competitive antagonist of dhfr. Gene amplification leads to more copies of the dhfr and TNF inhibitor genes present in the cells and, concomitantly, increased levels of TNF inhibitor mRNA which, in turn, leads to more TNF inhibitor protein being produced by the cells.

Example 10. Isolation of two types of TNF-inhibitors from U937 condition medium and the existence of the second TNF inhibitor in human urine.

The human U937 cells were grown at a density of 1 x 10^5 cells per ml in 150 cm^2 flasks using RPMI1640 medium containing 200 units/ml penicillin, 200 units/ml of streptomycin, 10% fetal calf serum. After 3 days of incubation at 37°C, the cells were harvested by centrifugation at 1500 G for 7 minutes. The cells were resuspended at a density of 2 x 10^6/ml in RMPI1640 medium without serum. The cells were grown in the presence of 5 ug/ml PHA-P (Phytohemagglutinin) and 10 ng/ml PMA (Phobol 12-myristate 13-acetate) for 24 hours.

The 24 hour medium (4425ml) was collected by centrifugation and concentrated by Amicon YM5 filter to about 100 ml. This material was passed through a TNF-affinity gel (0.7 x 2cm) at a flow rate of 0.1 ml/min and the gel was washed extensively with 50mM Tris-HCl pH 7.5. The bound proteins were eluted with 50 mM $NaPO_4$-HCl, pH 2.5 and TNF inhibitor was separated from other contaminating proteins by HPLC-RPC8. As seen in Figure 27 two TNF-inhibitor peaks are observed. SDS-PAGE analysis of the RPC8 fractions shows that the molecular weights of the two peaks correspond roughly to 30kDa and 40kDa proteins (Figure 28). The 30kDa protein (TNF-1NH1) was subjected to amino-terminal sequence analysis, and found to be the same sequence as that of urinary 30kDa TNF-inhibitor described above. However, the protein sequence of the 40kDa protein reveals that it is not the same as the 30kDa protein (see Example 11). Further purification of the second TNF inhibitor peak in the human urine, which is seen around fraction 35 in Figure 8, shows that it is also the 40kDa TNF-inhibitor protein (Figures 29 and 30).

The 40kDa TNF inhibitor is also a glycoprotein. This was detected using Concanavalin A-peroxidase after the

protein was transferred onto a nitrocellulose filter as described in Example 1.D. The molecular weight of N-glycanase treated 40kDa TNF inhibitor was shown on SDS-PAGE to be about 36kDa. (See procedure described Example 1.D.).

Following the procedures as outlined in Example 1.E. above, it may be determined that the deglycoslyated 40kDa TNF inhibitor also binds to TNF alpha. In addition, the deglycosylated 40kDa protein may also be shown to bind to TNF beta (lymphotoxin).

Example 11. Protein sequencing of U937 derived 30kDa TNF inhibitor, 40kDa TNF inhibitor, and Urinary 40kDa TNF-inhibitor.

Amino terminal sequence of the proteins were determined using Applied Biosystem Protein Sequencer, Model 470. Both native and reduced-carboxymethylated proteins were sequenced. Approximately 200 pmoles of reverse phase (RP-8) purified TNF inhibitors were applied to a polybrene filter and subjected to automated Edman degradation. The resulting sequence is shown in Figure 31. It can be seen that the U937-derived 30kDa protein is the same as that formed and identified in urine. The 40kDa TNF inhibitor protein is not the same as the 30kDa TNF inhibitor protein. The urinary 40kDa TNF inhibitor protein does not contain two amino terminal residues; otherwise, it is same as that of the U937-derived 40kDa protein.

Example 12. Primary structure of the 40kDa TNF inhibitor.

About 40 ug of the reduced and carboxymethylated TNF inhibitor (40kDa) was digested with endoprotease V8 as described above, and the resulting peptides were separated on an RPC18 column (Figure 32). The peptides purified were sequenced using an Applied Biosystem Protein Sequencer, Model 470.

About 90 ug of the reduced and carboxymethylated TNF inhibitor was treated with 5 ug of endopeptidase Arg-C in 0.2M ammonium bicarbonate at 37°C. After 24 hours of digestion, the Arg-C digested material was loaded onto an HPLC-RP8 column to separate peptides (Figure 33). Purified peptides were sequenced as before. Some of the peptides were further digested with TPCK-trypsin or chymotrypsin. About 500 pmole of arg-C16 peptide was treated with 3 ug of TPCK-trypsin (Boehringer Mannheim) in 0.2M ammonium bicarbonate at 37°C for 7 hours, and peptides were separated using RP8 (Figure 34). About 200 pmole of the peptide arg-C10 was digested with one ug of chymotrypsin (Boehringer Mannheim) at 37°C for three and a half hours, and the resulting peptides were separated on an RPC18 (Figure 35).

A partial structure of the TNF inhibitor (40kDa) was determined by aligning various overlapping peptides (Figure 36). A complete primary structure of the 40kDa TNF inhibitor is shown in Figure 38. Residues not identified by protein sequencing were deduced by review of the sequence of the cDNA clone that encodes the 40kDa TNF inhibitor and that is discussed in Example 14A and described in Figure 39.

Example 13. Identification of cDNA clones for the 40kDA TNFa inhibitor.

The information presented in Example 9 shows that U937 cells treated with PMA and PHA produce a TNFa inhibitor with a molecular weight of approximately 40kDa. This protein has been purified and it's amino acid sequence has been substantially determined, as described in Example 12. Table 5 shows the sequences of several peptides derived from this protein and gives the sequences of mixed sequence oligonucleotide probes used to isolate genes coding for the 40kDa TNF inhibitor described here.

The gene encoding sequences comprising the 40kDa

inhibitor may be isolated from the human genomic library described in Example 5, or a cDNA library constructed from mRNA obtained from U937 cells that had been treated with PMA and PHA for about 9 hours (See Example 14). Each library should contain approximately 1.0×10^6 recombinant.

TABLE 5

Peptide Sequence	Probe Name	Probe Sequence
EYYDQTA	40KD-P2'	5'GAATATTATGATCAAACAGC 3' C G C C C G G T
AQUAFT	40KD-P1	5'GTAAAACGAACTTGAGC 3' C C C G G G C G T T T
KQEGCR	40KD-PG	5'AAACAAGAAGGATGTCG 3' C G G G G CAC T
QMCCSKC	40KD-P5	5'CATTTAGAACAACACATTTG 3' C C GCTG G C T
DQTAQMC	40KD-P6'	5'GATCAAACAGCACAAATGTG 3' C C C G G G G T T
PGWYCA	40KDP7	5' CCAGGATGGTATTGTGC 3' C C C C G G T T

Example 14. Isolation of 40kDa TNF inhibitor cDNA sequences from PMA/PHA-induced U937 cells

U937 mRNA was isolated from cells that had been induced by PMA/PHA for 9 hours. It was then selected on an oligo-dT column, and the polyadenylated mRNA thus isolated was used to make dscDNA using reverse

transcriptase followed by E. coli polymerase I/RNase H. The dscDNA was subjected to a polymerase chain reaction using, as primers, the degenerate probes (40KD-P1' and 40KD-P7) shown in Table 5. The DNA products from this reaction were probed on a Southern blot with probe 40KD-P6' (see Table 5) identifying a single band that contained this sequence. This band was isolated on an agarose gel and cloned into M13 phage DNA (strain mp18). After transformation into E. coli strain JM109 and plating on medium containing X-gal and IPTG, clear plaques were identified that contained the correct cDNA insert. The sequence of the DNA in this clone is shown in Figure 37 along with the translation product predicted from this sequence. This amino acid sequence matches the peptide sequence shown in Figure 36 (residues 12 - 104) and Figure 38.

Example 14A. Isolation of 40kDa TNF inhibitor cDNA clone from PMA/PHA-induced U937 Cells

mRNA was isolated (Chirgwin, J.M. et al., Biochemistry 18, 5294-5299) from human U937 cells that had been exposed to PHA and PMA for 9 hours. mRNA was purified from this RNA using oligo-dT cellulose (Aviv, H. and Leder, P., 1972, Proc. Natl. Acad. Sci. (USA) 69, 1408-1412). 5 ug of this mRNA was used to synthesize 3 ug of blunt-ended, double-stranded cDNA (Gubler, U. and Hoffman, B.J., 1983, Gene 25, 263-269). After addition of EcoRI linkers, the cDNA was purified by sephacryl S-400 (Pharmacia) spun column chromatography and ethanol precipitated. One hundred ng of this cDNA was ligated into 1 ug of EcoRI-digested and alkaline phosphatase-treated lambda gt-10 and packaged in vitro using giga-pack gold (Stratagene). The packaged cDNA yielded 2.5 x 10^6 recombinants when plated on E. coli C600 hfl. 1.2 x 10^6 members of this library were screened in duplicate with ^{32}P-labeled probe 40KD-P6+7 (5' GGG CGT ATG TGC TGT CCT CAC AGG 3') as described (Benton, W.D. and Davis,

R.W., 1977, Science 196, 180-182). Twelve positive hybridizing clones were isolated and rescreened with probes 40KD-P6' and 40KD-P7 (see Table 5 in Example 13). Four of these clones hybridized to all three probes. One of these clones, c40DK#6, was digested with EcoRI, and a 2.2 kb insert was isolated and subcloned in both orientations into the bacteriophage M13 vector, mp19 (Yarrish-Perron, C., et al., 1985, Gene 33, 103-119). The sequence was determined from both strands using the chain termination method (Sanger, F. and Coulson, A.R., 1975, J. Mol. Biol. 94, 441-448) with Taq DNA polymerase (U.S. Biochemical). This sequence is shown in Figure 39 along with its deduced translation product. The sequence contains a single open reading frame extending from the ATG triplet at base 93 that extends well beyond the c-terminal sequence of the 40kDa protein at the GAC triplet at base 863.

Example 15. The 40kDa TNF inhibitor inhibits TNF beta as well as TNF alpha

Both the 30kDa TNF inhibitor and the 40kDa TNF inhibitor were examined to determine if they were also capable of inhibiting the activity of TNF beta (lymphotoxin). Various concentrations of TNF-beta (purchased from Endogen) were incubated with each of the inhibitors for one hour at room temperature. The resultant mixtures were analyzed via the L929 cell assay system as described in Example 1.B.1. for TNF alpha. These experiments revealed that the 30kDa TNF inhibitor had little inhibitory effect on TNF beta. However, the 40kDa TNF inhibitor showed significant TNF beta inhibition. The results of these experiments can be seen in Figure 40.

Example 16. Preparation of human genomic DNA library for 40kDa inhibitor

An appropriate human genomic DNA library for 40kDa

TNF inhibitor may be performed as described in Example 5 for 30kDa TNF inhibitor.

Example 17. Preparation of genes for the Expression of the 40kDa TNF inhibitor cDNA in Escherichia coli

Portions of the TNF inhibitor (40 kDa) cDNA gene coding for soluble TNF binding activities (Fig. 39) have been prepared for expression in E. coli as described below.

Because it has been difficult to definitively determine the C-terminal sequence of the mature 40kDa TNF inhibitor derived from urine or U937 cells, we constructed 3 derivatives of its cDNA coding sequence based on sequence analysis of the cDNA clone. The first extends to the putative transmembrane sequence of this protein base pair 863 (Figure 39) and ends with the peptide sequence . . . Gly Ser Thr Gly Asp. The next two are 51 (Δ51) and 53 (Δ53) amino acids shorter than this clone and end at base pair 710 . . . Ser Pro Thr, and base pair 704 . . . Ser Thr Ser, respectively.

Each of these three C-termini were created by in vitro mutagenesis ("MutaGene", BioRad, Richmond, CA) of M13 clones of the cDNA of the 40 kDa TNFa inhibitor. The longest clone was created first by use of the following synthetic oligonucleotides:

1. 5' CAC TGG CGA CTA AGC TTC GCT CTT C 3'
2. 5' GCG GCG CAC GCC GGA TCC GAT CTT GGA GGA TGA TTA AAT GTT GCC CGC CCA G 3'

Oligonucleotide 1 inserts a translation termination codon after amino acid 235, Asp, and creates a HindIII restriction endonuclease recognition site at that point. Oligonucleotide 2 adapts the N-Terminal sequence of the mature protein, Leu Pro Ala . . . bp 159 (Figure 39) for expression in E. coli by 1) inserting a Met, ATG codon at amino acid position 1, and 2) inserting a translational coupler sequence and 5' BamHI restriction endonuclease recognition site. The mutagenized fragment was removed

by BamHI/HindIII digestion of Rf DNA of the mutant M13 clone and inserted into an E. coli expression plasmid as described in Example 7. Clones bearing this gene construction are called TNF.40.

The two shortened clones were constructed as above using the mutagenized M13 derivative of the 40kDa TNFa inhibitor clone isolated above and the following oligonucleotides:

5' GTCCCCCACCTAAGCTTCGGAGTATGG 3' Δ51

5' GTCCACGTCCTAAGCTTCCCACCCGGA 3' Δ53

These two oligonucleotides introduce translation termination codons at bp 710 and 704 respectively (Figure 39). Clones bearing these gene constructions are called TNF:40 Δ51 and TNF: 40 Δ53 respectively.

Example 18. Expression of genes encoding 40kDa TNF inhibitor in animal cells

Expression of the 40kDa TNF inhibitor clone in animal cells may be performed as described in Example 9. The extensive region located 3' of the c-terminus of the 40kDa TNF inhibitor may be deleted and a stop codon engineered into the position just following the c-terminal Aspartic acid.

Example 19. Expression of the complete cDNA encoding 30kDa TNF inhibitor in mammalian cells increases TNF receptor sites

An expression vector was made that incorporated the entire 30kDa TNF inhibitor cDNA (2.1 kb) shown in Figure 21, named p30KXVA, and was in all other respects identical to the vector shown in Figure 23 (i.e., the TNF-BP sequences shown in that figure were replaced by the 2.1 kb cDNA using the unique EcoRI site in the plasmid). See Example 9 for a more complete description of the expression vector. This plasmid was introduced into COS7 cells using the lipofection procedure described

by Feigner et al. (Proc. Natl. Acad. Sci. USA, 84, 7413 (1987)). Transfected cells were analyzed for their ability to bind [^{125}I]TNFa. Figure 41 shows the results of the binding assay of cells that were mock-transfected or transfected with the expression vector p30KXVA. The number of binding sites on plasmid-transfected cells is dramatically higher that the number on the control cells. The complete cDNA clone (i.e., the open reading frame that encodes a much larger protein than the 30kDa urine-derived inhibitor), in fact, represents a cDNA clone of a TNF receptor.

Example 20. Expression of cDNA encoding 40KDa TNF inhibitor in mammalian cells increases TNF receptor sites

An expression vector was made using the 2.4 kb cDNA fragment isolated from the lambda phage page #6 described in Example 14A. This plasmid was identical to that described in Example 9 (Figure 23) except that the 40kDa TNF inhibitor cDNA sequences were substituted for the 30kDa TNF inhibitor cDNA sequences in that plasmid. Plasmids were isolated with the 2.4 kb EcoRI cDNA fragment in each orientation, named p40KXVA (sense orientation) and p40KXVB (anti-sense orientation). These plasmids contain the SV40 origin of replication, the cytomegalovirus immediate early promoter and enhancer, the rabbit B-globin second intron, the 40KDa TNF inhibitor cDNA, and the SV50 early polyadenylation signal (for a more complete description of this vector, see Example 9) in a pBR322-based plasmid. These plasmids were transfected into COS7 cells which were then assayed for TNF binding (see Figure 42). Cells transfected with p40KXVA exhibited a higher number of TNF binding sites on the cell surface than either COS7 cells alone or COS7 cells transfected with p40KXVB, suggesting that this cDNA encodes a TNF receptor. Other mammalian cells such as CHO cells could be developed that could overproduce this receptor or that secrete 40KDa TNF inhibitor into the

tissue culture medium in ways described in Example 9.

Example 21. Inhibitor isolated from human monocytes.

Human monocytes were prepared from 550 ml of blood as described by (Hannum, C.H. et al. Nature 343, 336-340, 1990). The fresh monocytes (2 x 10^7 cells) were seeded in 500 ml of serum free RPMI1640 medium and treated with 10ng/ml of PMA and 5ug/ml of PHA-P for 24, 48 and 72 hours at 37°C. After the incubation, the media were collected by centrifugation and concentrated to 50 ml. The concentrated media were loaded onto a TNF-affinity column (2 ml bed volume) one sample at a time and eluted with acid as in Example 1. The eluted material was further purified using a HPLC RPC-8 column under the same conditions as in Example 1, and each fraction was assayed with L929 cytoxicity assay. Figure 43 shows the two peaks of TNF inhibition activity. These two peaks correspond to the 30 kDa and 40 kDa TNF inhibitors which were also found in the culture medium of U937 cells that was treated with PMA and PHA and identified in urine.

Example 22. Expression and Purification of shorter forms of the 40 kDa TNF inhibitor (Δ51 and Δ53) from E. coli.

Cells (300 ml of E. coli cultures (40kDa TNF inhibitor Δ51 and 40kDa TNF inhibitor Δ53)) grown separately under induced condition for 2 hours were resuspended in 10 ml of 50 mM Tris-HCl, pH 7.5 containing 2mM EDTA (TE buffer) and French pressed at 20,000G for 10 min. The resulting pellets were washed once with TE buffer. The washed pellet was resuspended in 2 ml of 6M Guanidine-HCl/100mM Tris-HCl, pH8.5/4mM PMSF, and incubated at room temperature for one hour. After incubation, 500mM DTT was added to a final concentration of 4mM, and the mixture was incubated at room temperature for another hour. Insoluble material was removed by centrifugation at 20,000G for 15 min. 500mM oxidized glutathione was added to the supernatant to a final

concentration of 20mM, and the mixture was incubated at room temperature for 10 min. This material was then diluted in 20 ml of 0.6% Tris base solution with 5mM cysteine. PMSF was added to a final concentration of 2mM. After 16 hours of incubation at 4°C, this material was dialyzed against 300 volumes of 50mM Tris-HCl, pH 7.5 for 3 hours at 4°C, then centrifuged at 20,000G for 15 min. The supernatant was loaded onto a TNF-affinity column (.7 x 2 cm, 13 mg rhTNF/ml of affigel-10) at a flow rate of 0.09 ml per min. This column was extensively washed with 50mM Tris-HCl, pH 7.5. The bound proteins were eluted with 50mM NaH2P04-HCl, pH 2.5. The acidic eluates were loaded onto an RP8 column (2 x 200mm, spelco) and the TNF inhibitors were eluted with a linear gradient of acetonitrile in 0.1% TFA at a flow rate of 1 ml per gradient per min. (Figure 44A and 45A). Fractions were examined by L929 cytotoxicity assay to localize the TNF inhibitors. The major peak on each RP8 profile contains the TNF-inhibiting activity (Figure 44C and 45C). The E. coli-produced TNF-inhibitors (40kDa TNF inhibitor Δ53 and 40kDa TNF inhibitor Δ51) migrate to the expected location on SDS-PAGE (Figure 44B and 45B). The amino terminal sequence of these materials shows that the E. coli-produced TNF-inhibitors have the following sequence:

Met-Leu-Pro-Ala-Gln-Val-Ala-Phe-Thr-Pro-Tyr-Ala-Pro-Glu

By using this procedure, about 150ug of each 40kDa TNF inhibitor (Δ51 and Δ53) was obtained from 30 ml of the culture. The yield was a few percent, however, the yield can be increased to over 30% by improving each step of this purification.

Both of these 40kDa TNF inhibitors (Δ51 and Δ53) inhibit not only TNF-alpha but also TNF-beta.

Example 23. Expression and Purification of full length 40kDa TNF inhibitor.

An active 40kDa TNF inhibitor was purified from an

E. coli strain carrying plasmids which have a gene for full length mature 40kDa TNF inhibitor (as in Example 12). The method used to isolate an active inhibitor was the same as that of example 22. This active inhibitor inhibits both TNF-alpha and TNF-beta, and the amino terminal sequence is same as shown in Example 22.

Example 24. Amino acid composition of the 40kDa TNF Inhibitor.

U937-produced mature 40kDa TNF inhibitor was analyzed for total amino acid composition by the PTC-amino acid analysis system. The actual and predicted composition data for full length mature 40kDa TNF inhibitor as shown in Figure 38 are shown in Table 6.

Example 25. Production of chemically modified TNF inhibitors.

In order to increase the half-life of the TNF inhibitors in plasma, TNF inhibitors which are chemically modified with polyethylene glycol (PEG) may be made. The modification may be done by cross linking PEG to a cysteine residue of the TNF inhibitor molecules. Since all of the cysteine residues in the TNF inhibitors form disulfide bonds, mutant TNF inhibitors may be constructed which contain an extra cysteine residue at the amino terminus, glycosylation sites, and the carboxyl terminus of each inhibitor. The mutagenesis may be carried out by PCR using oligonucleotides containing the desired mutation. As for the 30kDa TNF inhibitor, an extra cysteine residue was added at residue number 1, 14 or 105. These mutant proteins were expressed in E. coli by using the same system described in Examples 7, 22 and 23, and refolded to active TNF inhibitor. The mutant proteins are as active as the non-mutated proteins. Pegylation of these proteins will be carried out, and the activity will be assessed. The 40kDa mutants will be constructed as above and pegylation will be performed to

obtain active proteins and will have increased the stability of the TNF inhibitor.

It is to be understood that the application of the teachings of the present invention to a specific expression system will be within the capabilities of one having ordinary skill in the art in light of the teachings contained herein. Thus, it will be apparent to those of ordinary skill in the art that various modifications and variations can be made in the processes and products of the present invention. It is intended that the present invention covers these modifications and variations provided they come within the scope of the appended claims and their equivalents.

TABLE 6

	Calculated # by DNA sequence	Experimental #
Asx	14	13.0
Glx	23	22.6
Ser	25	23.2
Gly	14	17.8
His	4	4.5
Thr	26	23.9
Ala	17	17
Arg	14	15.1
Pro	26	22.3
Val	13	8.7
Ile	4	3.4
Leu	10	8.6
Phe	5	4.6
Lys	6	5.4
Tyr	5	5.0
Trp	3	ND
Met	3	ND
Cys	22	ND

ND: not determined.